

# From qualitative to quantitative formal methods for biochemical signalling pathways

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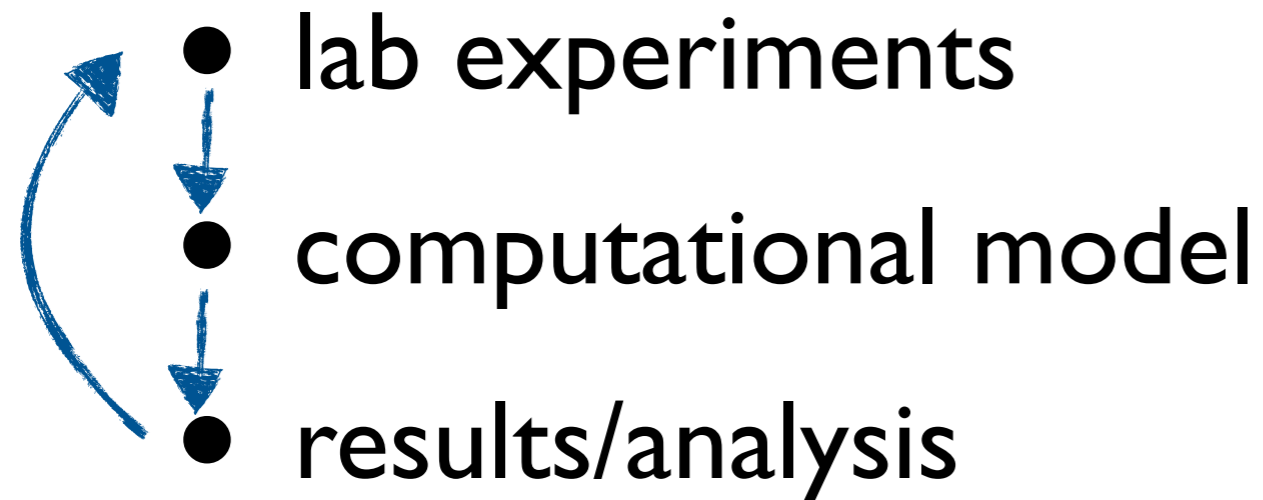
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joint work with H el ene Kirchner (INRIA Bordeaux) and  
Muffy Calder (University of Glasgow)

# Outline

- Motivation
- Rule-based modelling
- Abstractions for CTMCs
- Conclusion and perspectives

# Formal methods for modelling biological systems



Goals: to understand, to predict, to control

# Cell signalling

- communication between cells
- cellular processes: cell growth, proliferation, apoptosis...
- malfunctions may lead to diseases

# Challenges

- suitable formalisms
- abstraction techniques
- analysis
- scalability

# Our approaches

- qualitative: rule-based, higher-order calculus, runtime-verification
- quantitative: abstraction for CTMCs - CTMCs with levels, stochastic model checking

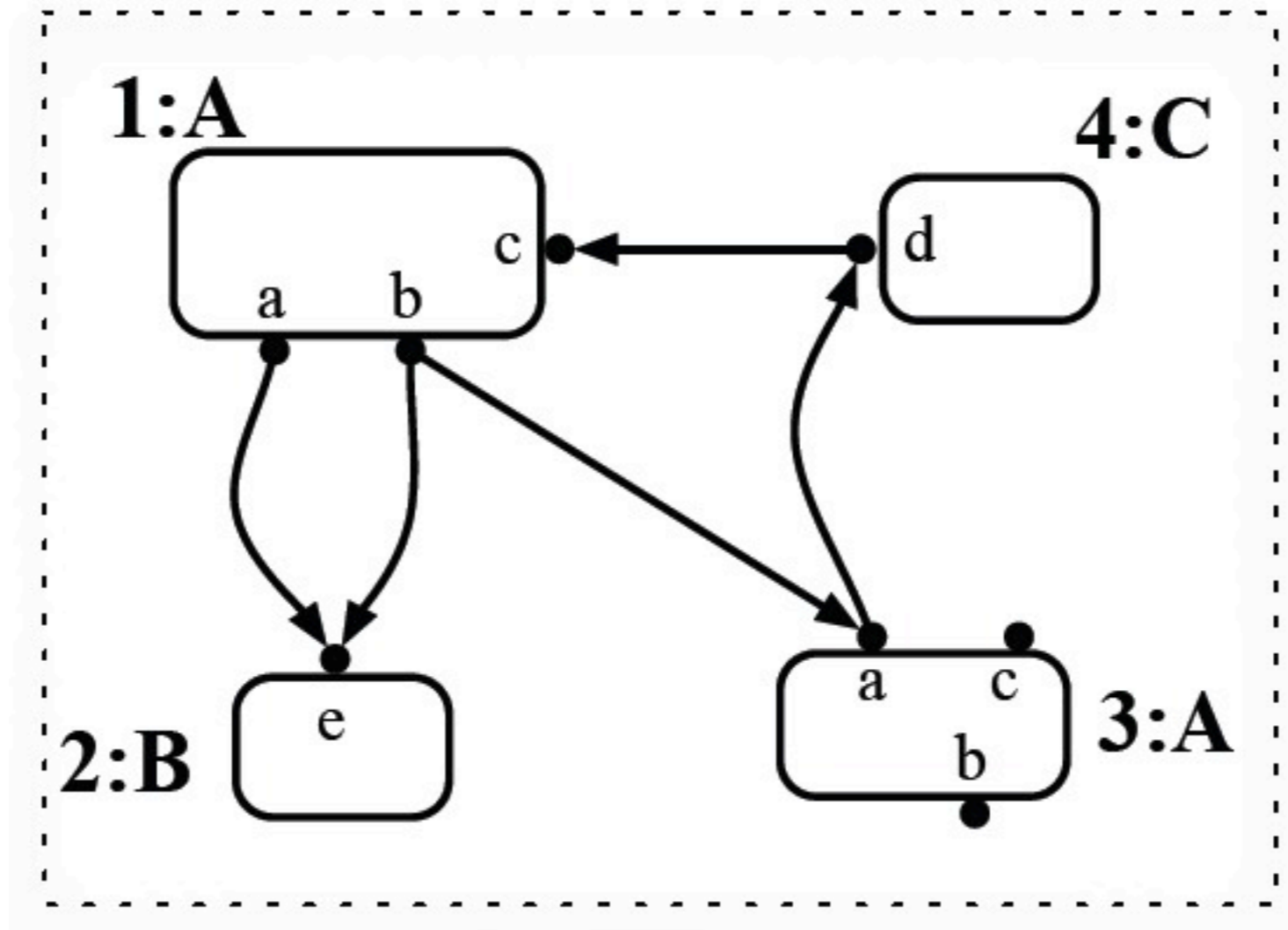
# Higher-order rule-based modelling

# Port graphs

- graphs with multiple edges and loops
- edges connect to **ports** of nodes
- defined over a signature  $(N, P)$



# A port graph



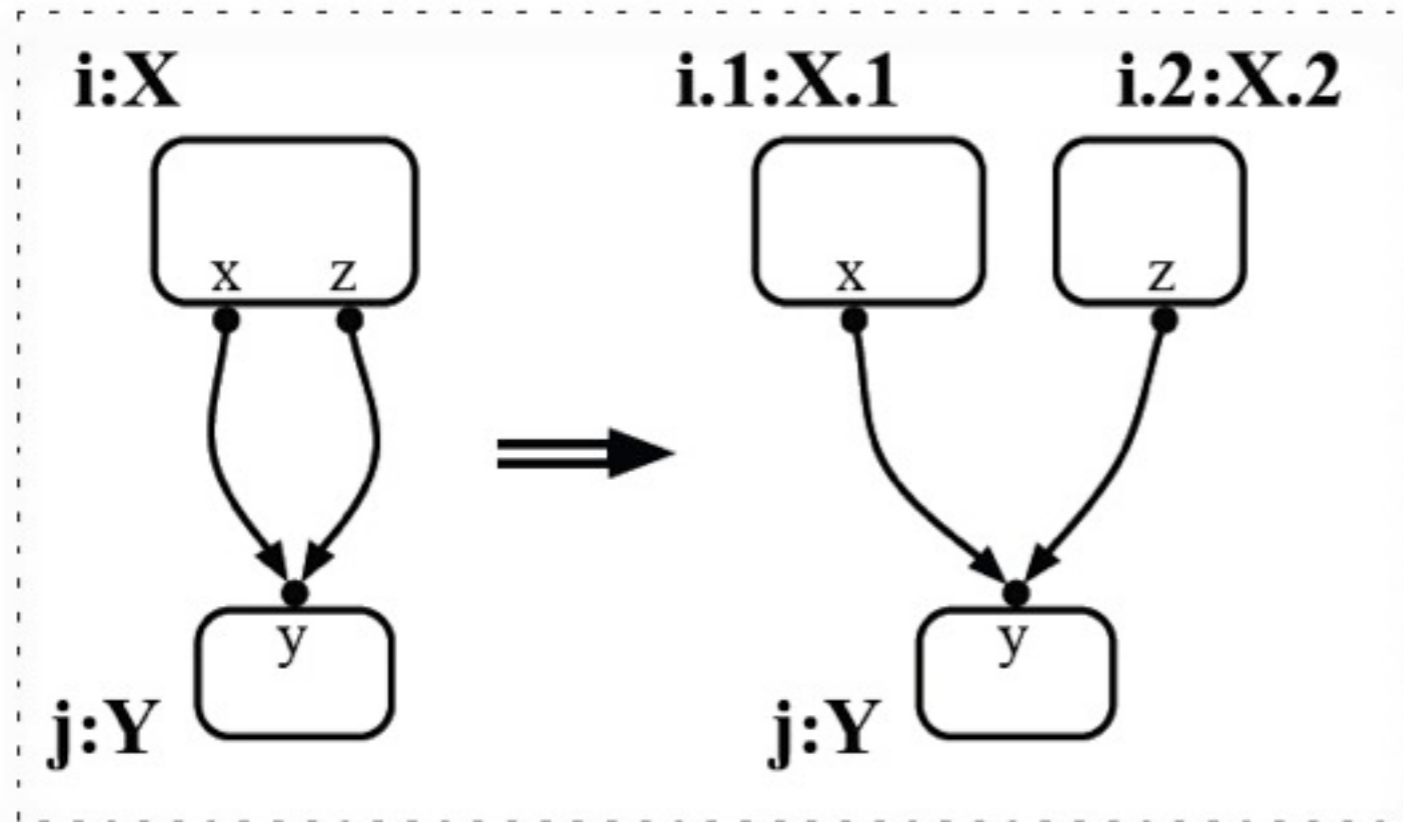
# Molecular graphs as port graphs

Molecular complex	Port graph
protein	node
site	port
bond	edge
interaction	rewrite rule

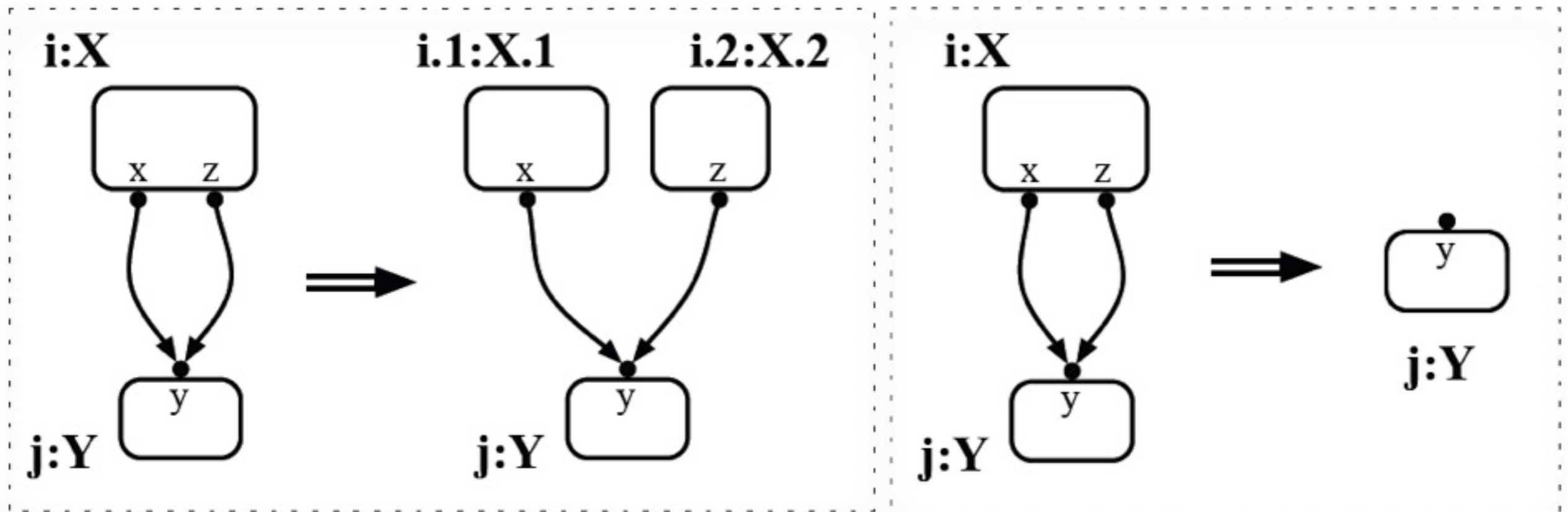
# Rewrite rules

- well-suited for modelling bio-molecular interactions
- a rule  $L \rightarrow R$  defines a class of reactions

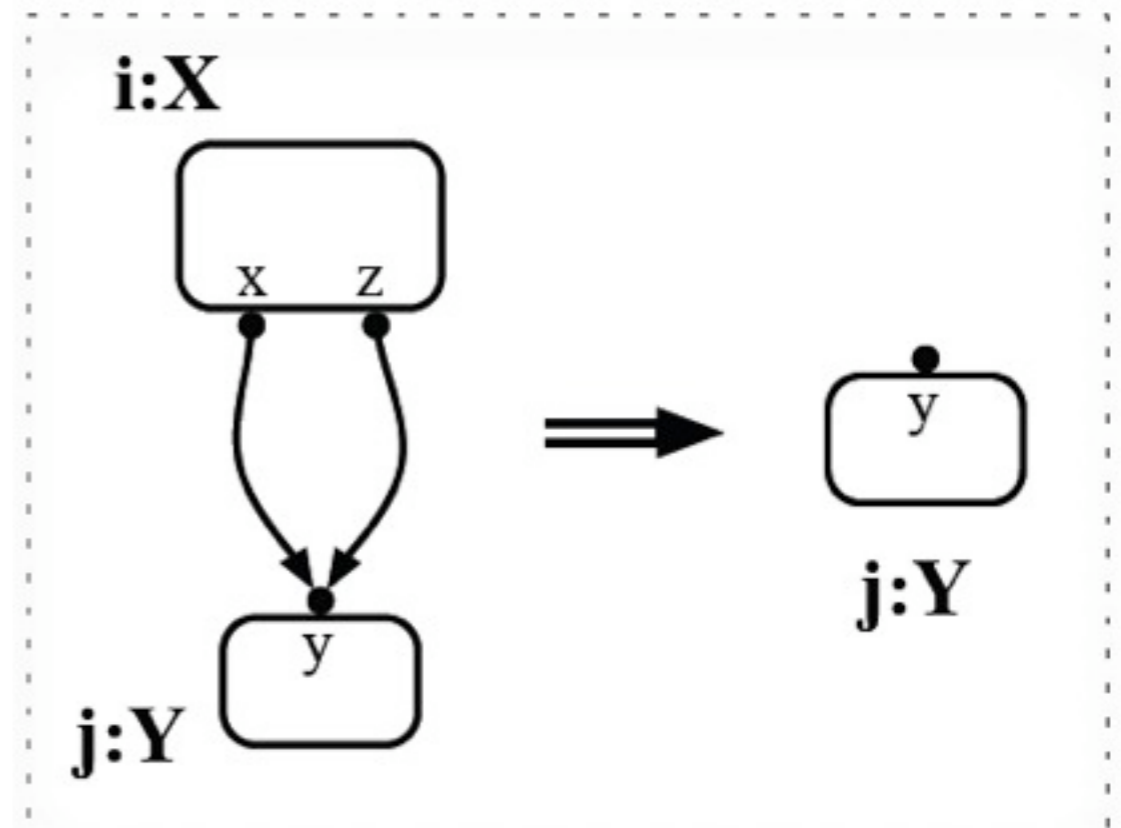
# Port graph rewrite rules



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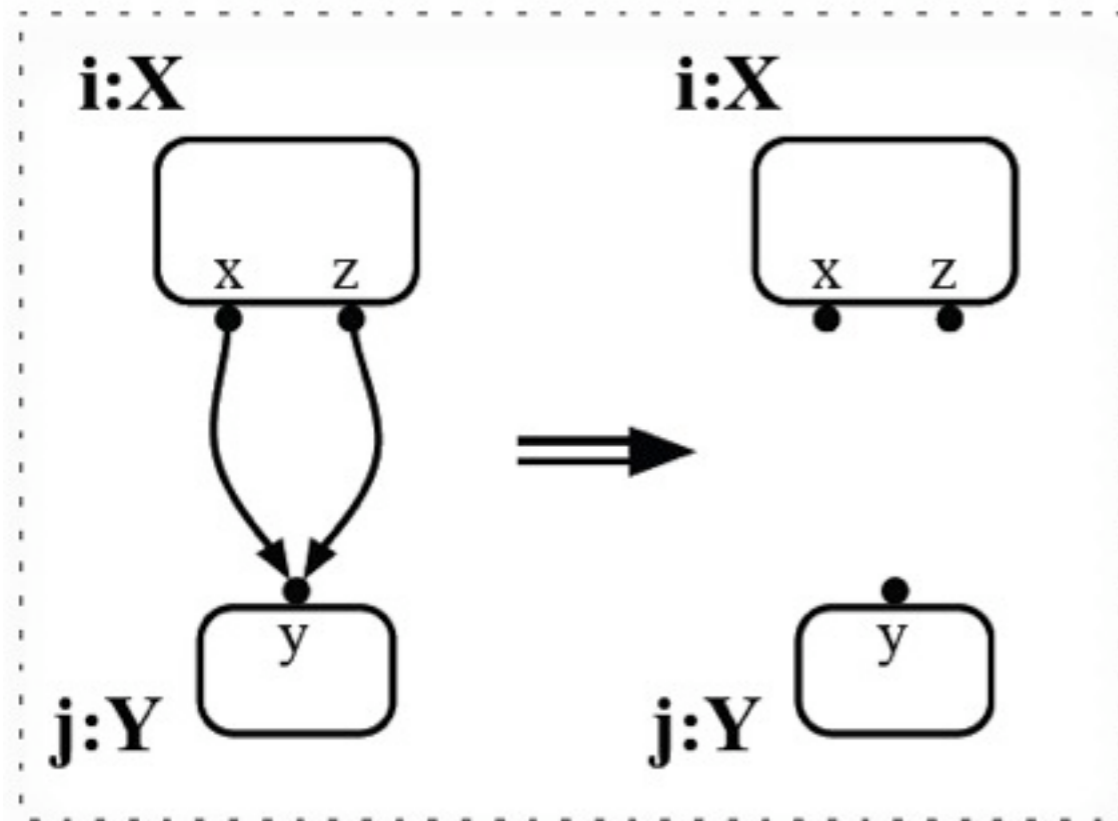


# Port graph rewrite rules



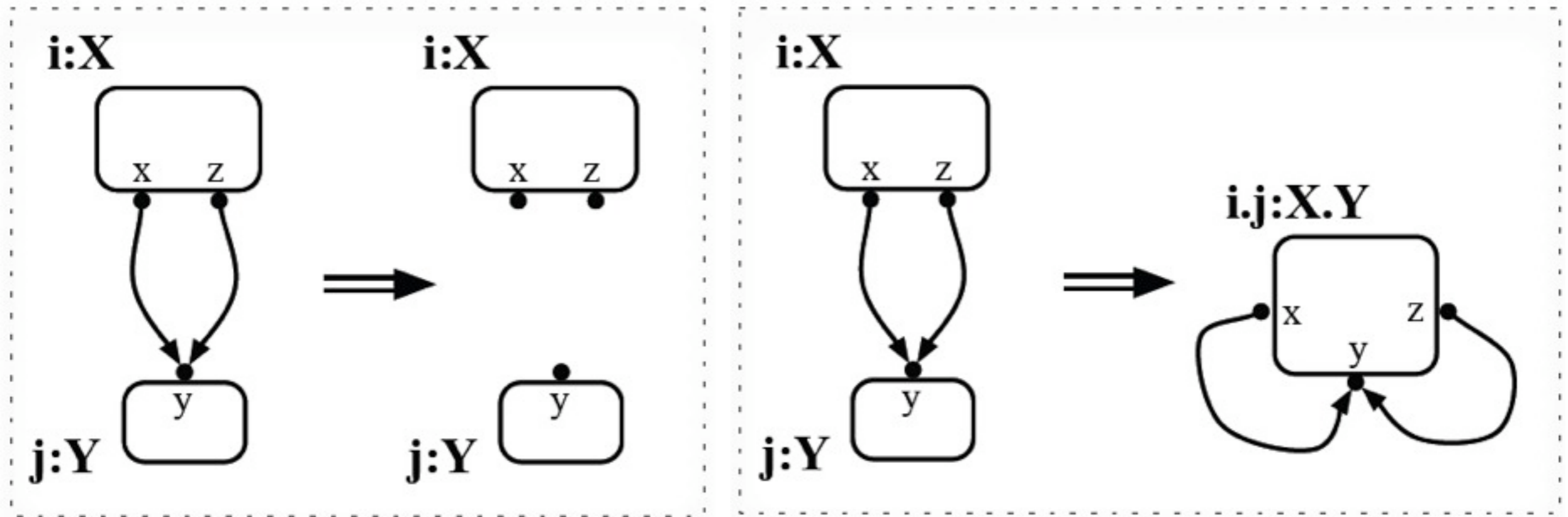
# Port graph rewrite rules

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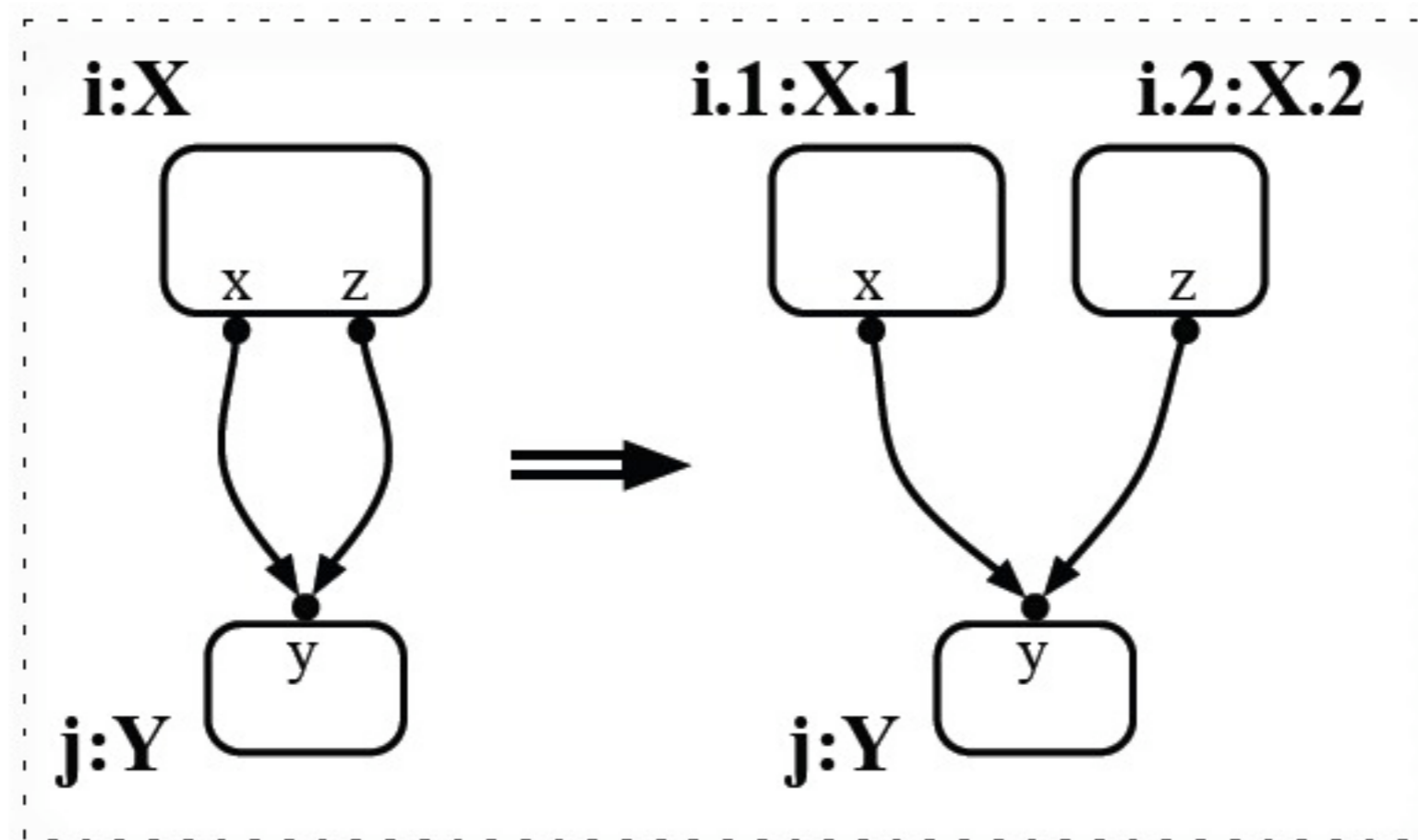




# Port graph rewrite rules

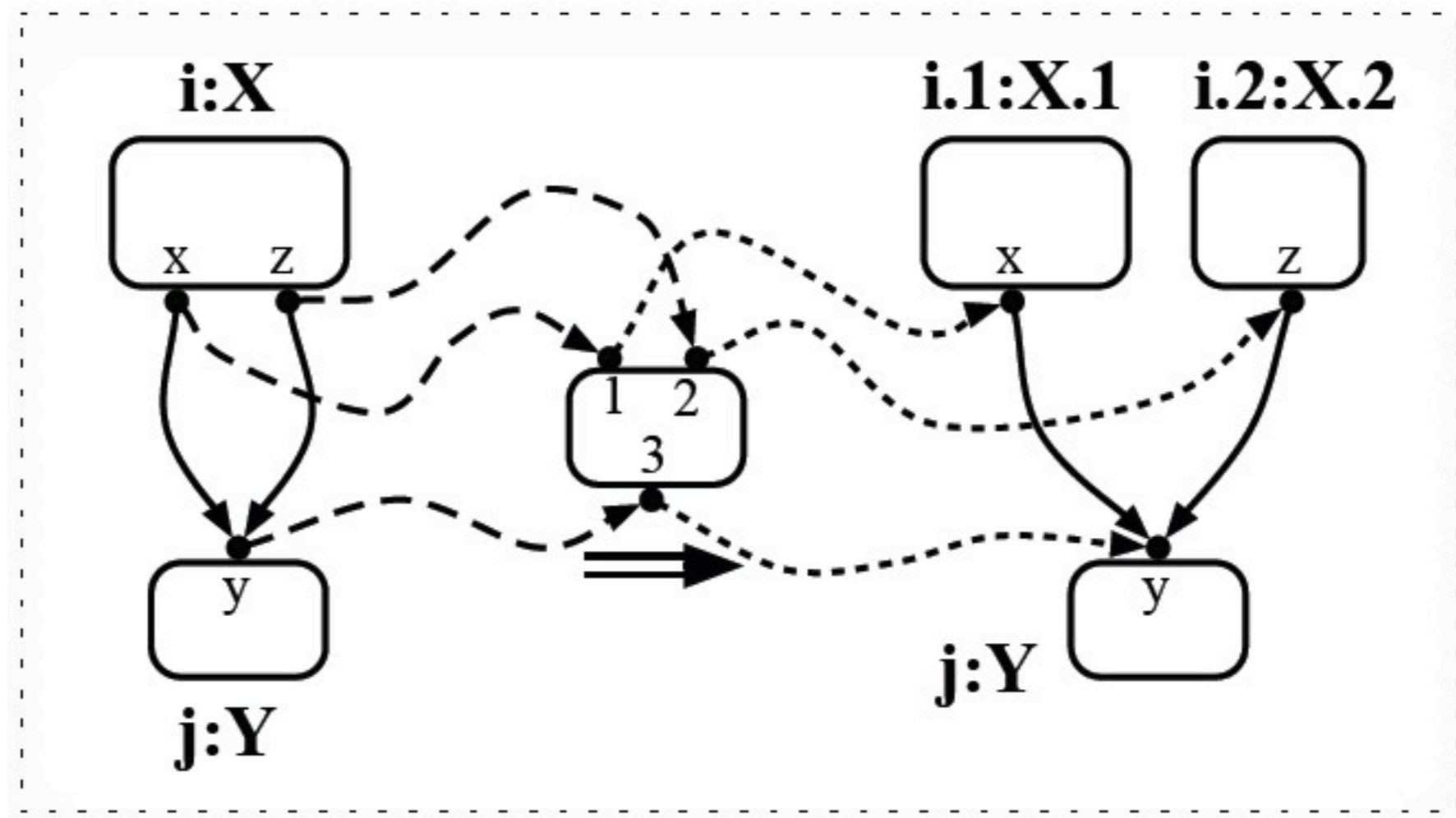


# A port graph rewrite rule is a port graph



A port graph rewrite rule is  
a port graph

# A port graph rewrite rule is a port graph



# Port graph rewriting relation

$$G \Rightarrow_{L \Rightarrow R} G' \quad \text{if} \quad \exists (g, G^-, \mathcal{B}) \in \text{Sol}(L \ll G)$$

such that

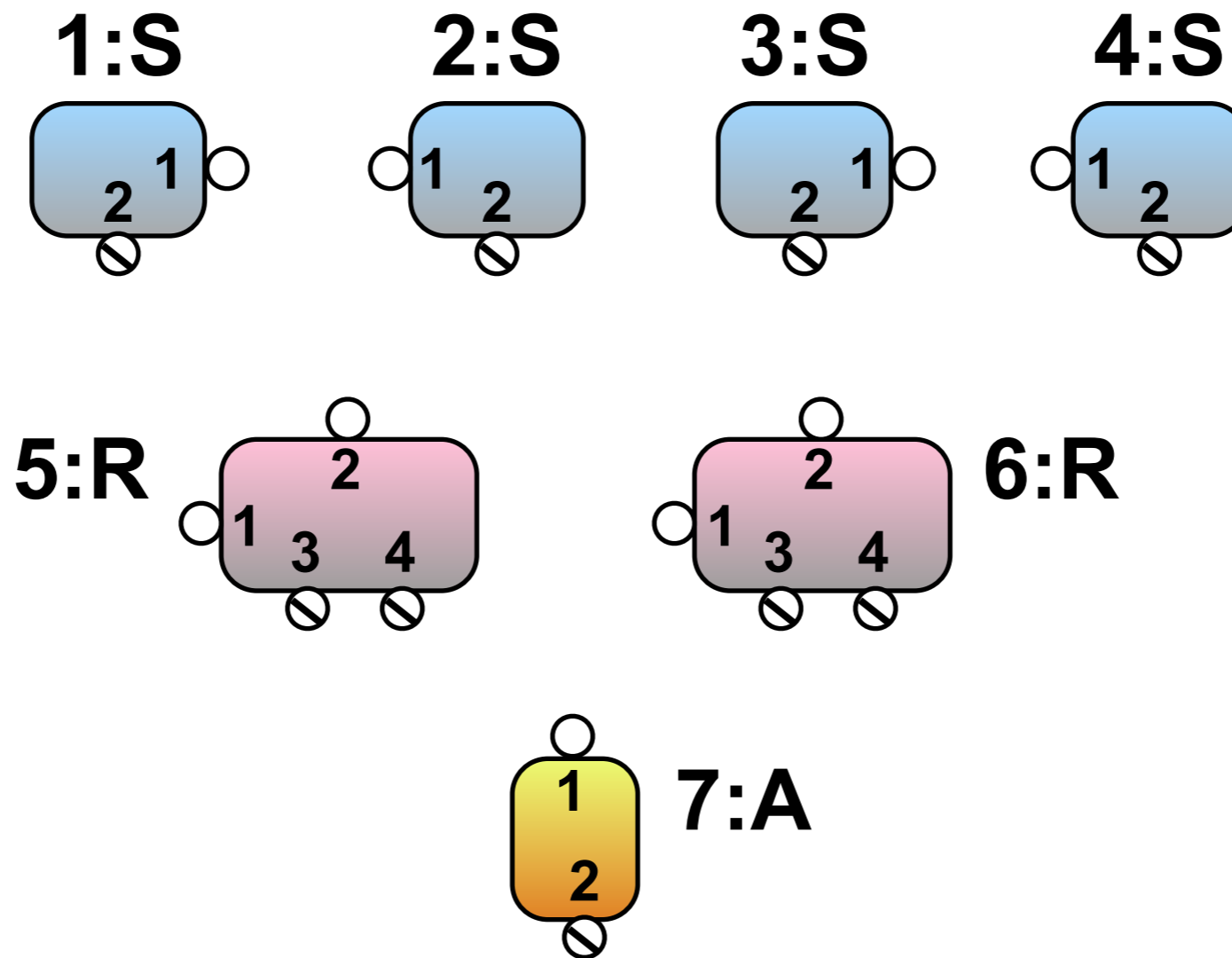
$$G = G^- [g(L)]_{\mathcal{B}}$$

and

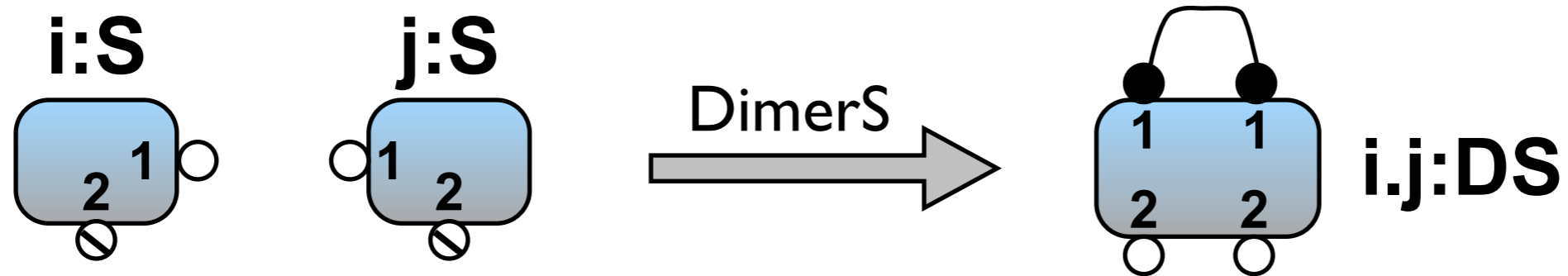
$$G' = G^- [g(R)]_{\downarrow_g \mathcal{B}}$$

# Example: a fragment of the EGFR signaling pathway

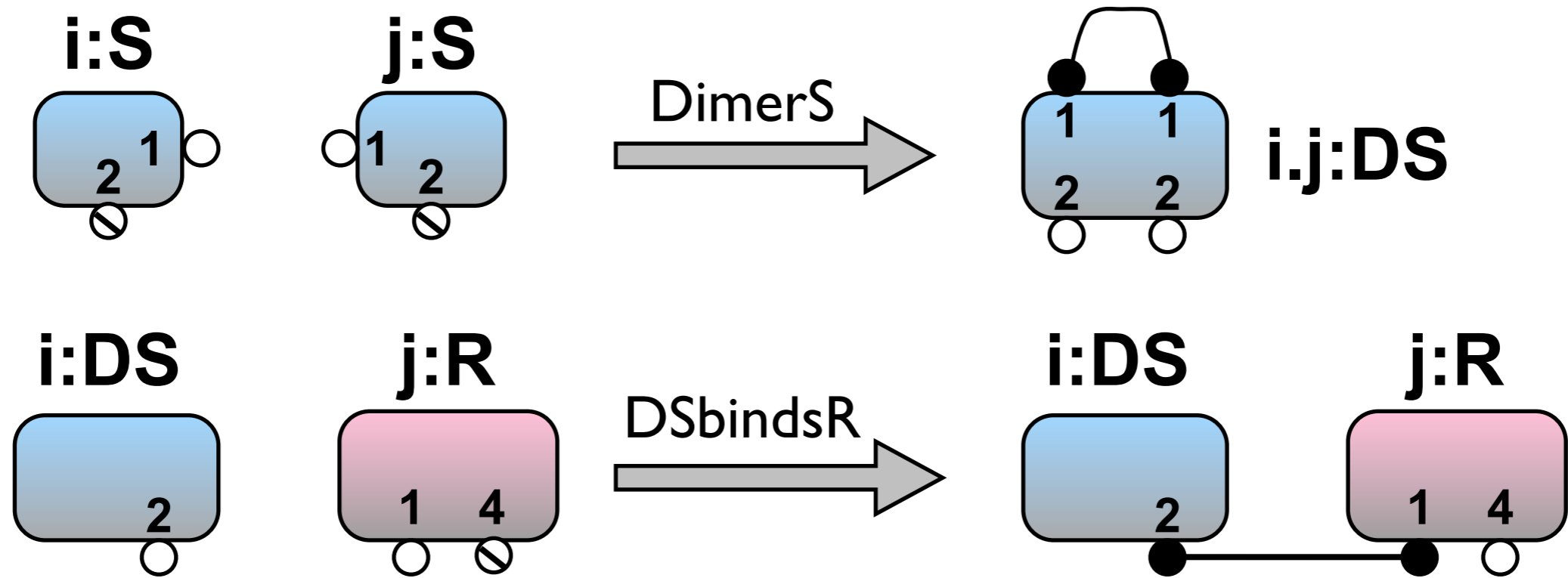
*Initial state:*



# Example: a fragment of the EGFR signaling pathway

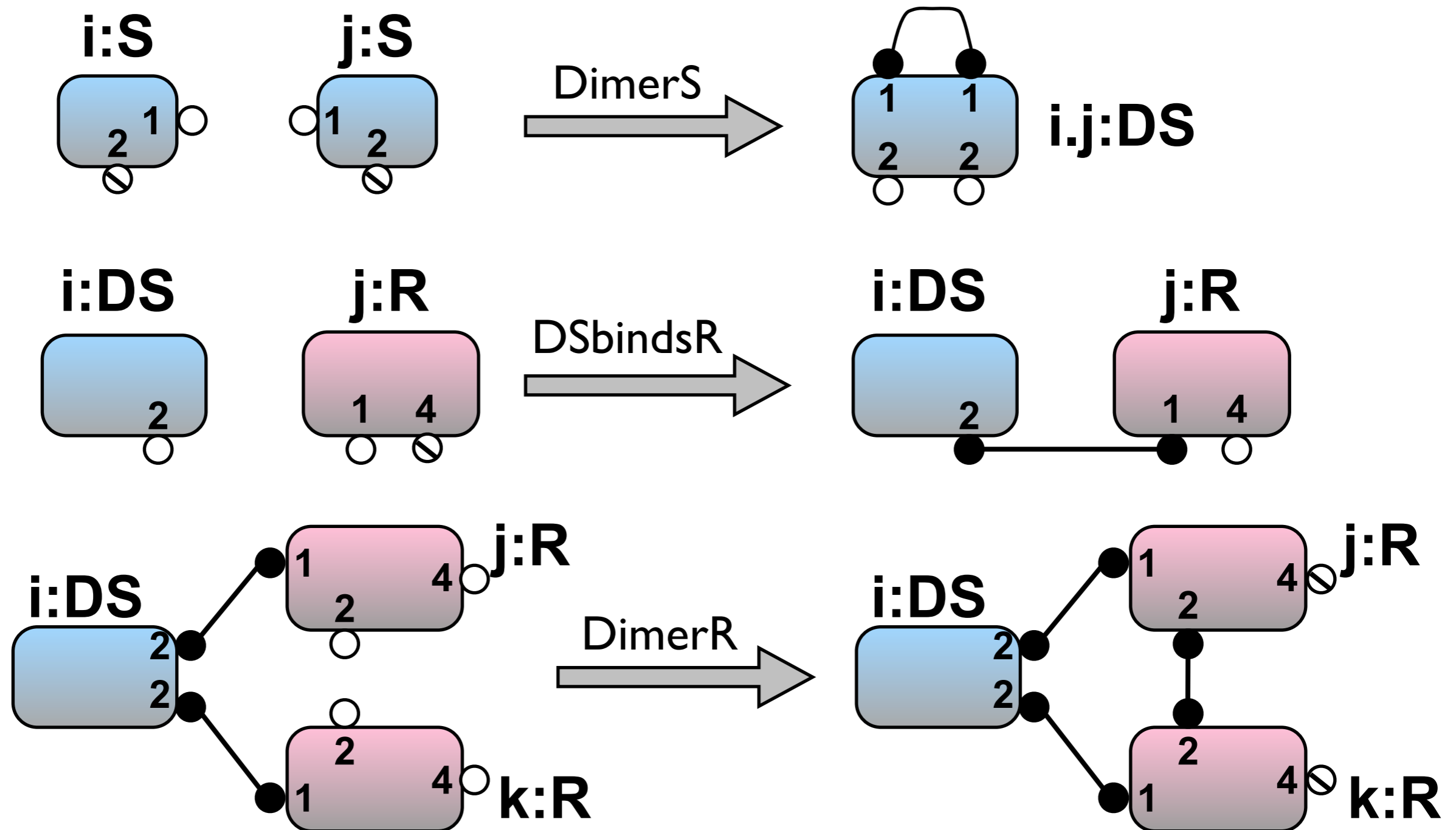


# Example: a fragment of the EGFR signaling pathway



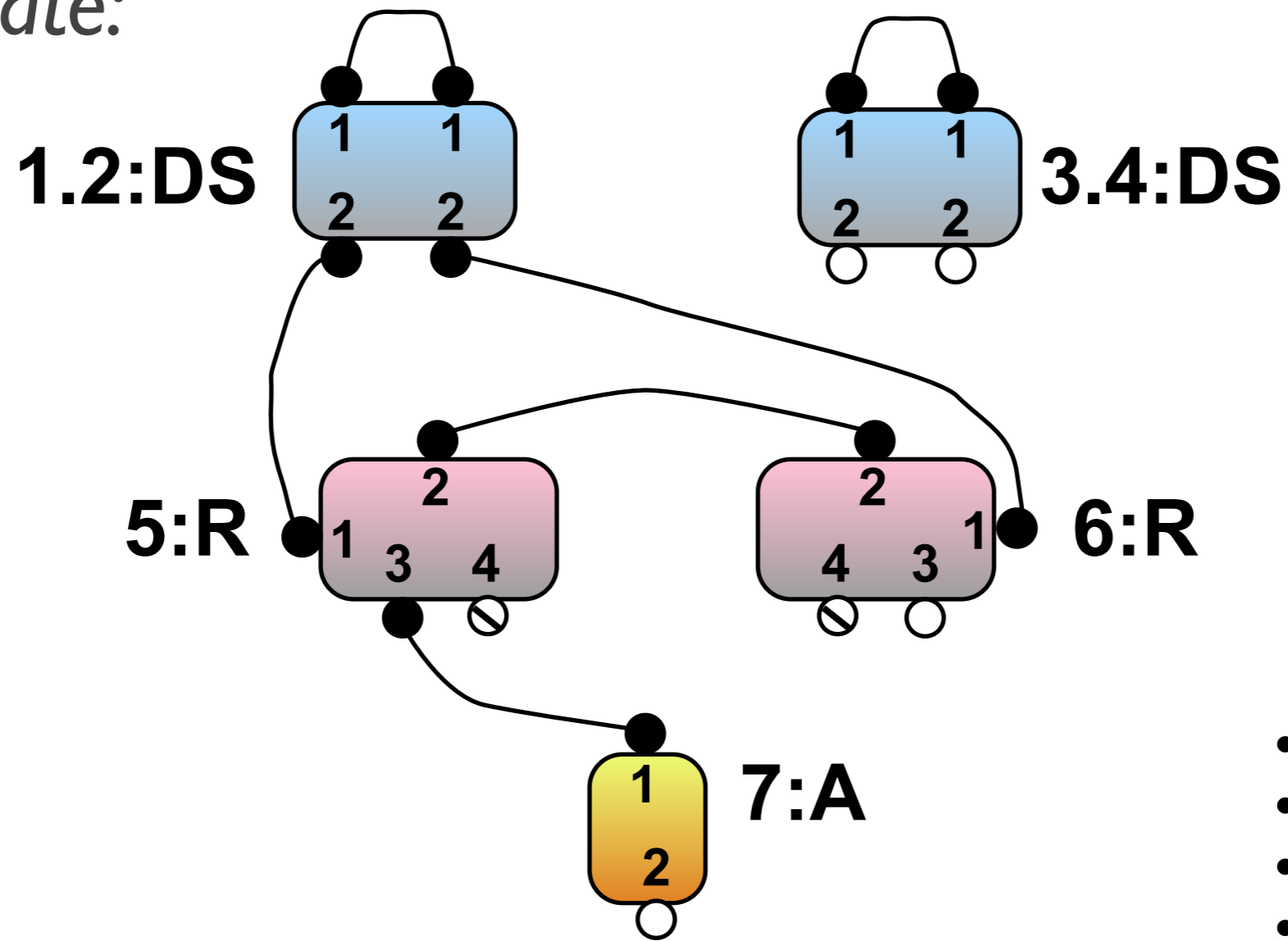


# Example: a fragment of the EGFR signaling pathway



# Example: a fragment of the EGFR signaling pathway

*A stable state:*



- 2 x DimerS
- 2 x DSbindsR
- 1 x DimerR
- 2 x ActivateDR
- 1 x DRbindsA

# Graph-base approaches

- $\kappa$ -calculus, Kappa factory [Danos et al.]
- BioNetGen [Hlavacek et al.]
- Pathway Logic [Talcott et al.]

# Chemical programming

- $\gamma$ -calculus =  $\lambda$ -calculus + chemical paradigm  
[BanatreFR04-07]
- a chemical solution where molecules interact freely according to reaction rules
- everything is a molecule

*prod* = replace  $X, Y$  by  $X \times Y$

$\langle \textit{prod}, 3, 1, 4, 5, 2 \rangle \rightarrow \langle \textit{prod}, 1, 4, 15, 2 \rangle \rightarrow^* \langle \textit{prod}, 120 \rangle$

# Rewriting calculus

- extends first-order term rewriting and the  $\lambda$ -calculus [CirsteaK01]
- terms, rules, rule application are explicit objects of the calculus

$$(s(x)+y \rightarrow s(x+y)) (s(5)+s(2)) \rightarrow_{\rho} s(5+s(2))$$

# Biochemical calculus

- add biochemical flavour to the chemical calculus - structures (like port graphs)
- rewrite strategies for controlling the rule application (*Identity, Failure, Sequence, Not, First, ...*)
- verification techniques

# Syntax

- objects: port graphs
- rewrite rules
- abstractions
- application

(Objects)  $\mathcal{O} ::= OBJ \mid \mathcal{X} \mid \mathcal{O} \bullet \mathcal{O}$

(Rule)  $\mathcal{R} ::= \mathcal{O} \Rightarrow \mathcal{O}$

(Molecule)  $\mathcal{M} ::= \mathcal{O} \mid \mathcal{R} \mid \mathcal{M} \bullet \mathcal{M}$

(Abstraction)  $\mathcal{A} ::= \mathcal{M} \Rrightarrow \mathcal{M}$

(Configuration)  $\mathcal{K} ::= \mathcal{M} \mid \mathcal{A} \mid \mathcal{K} \bullet \mathcal{K}$

(System)  $\mathcal{S} ::= [\mathcal{K}]$

# Semantics

(Interaction)  $[K \bullet (M \Rightarrow N) \bullet M'] \longrightarrow_i [K \bullet_{\varsigma}(N)]$  if  $\varsigma \in \text{Sol}(M \Leftarrow M')$



# More control? Use strategies

- provide control over the composition or choice of the abstraction to apply

- enforce confluence and termination

★ *Identity, Failure, Sequence, Not, First, Repeat...*

$First(S_1, S_2)(G) = S_1(G)$  if  $S_1$  does not fail,  
 $S_2(G)$  otherwise

- encoded as abstractions in the calculus

# Strategies-based extensions

- tackling application failure

(InteractionR)  $[K \bullet T \bullet M] \longrightarrow_{ir} [K \bullet \text{seq}(T, \text{try}(\text{stk} \Rightarrow T \bullet M)) @ M]$

◆ persistent strategies  $S!$

# Invariant verification

- invariant:
  - rule  $G \Rightarrow G$
  - strategy `first( $G \Rightarrow G$ ,  $X \Rightarrow \text{"Failure"}$ )!`
- remove `( $G \Rightarrow \text{"Failure"}$ )!` or “repair” `( $G \Rightarrow H$ )!`
- but we can do more...

# Structural formulas

# Structural formulas

Structural formulas:

$$\varphi ::= \top \mid \perp \mid \gamma \mid \neg\varphi \mid \varphi_1 \wedge \varphi_2 \mid \varphi_1 \vee \varphi_2 \mid \varphi_1 \rightarrow \varphi_2 \mid \diamond\varphi$$

# Structural formulas

## Structural formulas:

$$\varphi ::= \top \mid \perp \mid \gamma \mid \neg\varphi \mid \varphi_1 \wedge \varphi_2 \mid \varphi_1 \vee \varphi_2 \mid \varphi_1 \rightarrow \varphi_2 \mid \diamond\varphi$$

## Satisfaction relation:

$$G \models \gamma \quad \Leftrightarrow \quad \exists \sigma \text{ such that } G = \sigma(\gamma)$$

$$G \models \diamond\varphi \quad \Leftrightarrow \quad \exists G' \sqsubseteq G \text{ such that } G' \models \varphi$$

# Structural formulas as strategies

$$\begin{aligned}\tau(\top) &= \text{id} \\ \tau(\perp) &= \text{fail} \\ \tau(\diamond\gamma) &= \gamma \Rightarrow \gamma \\ \tau(\neg\varphi) &= \text{not}(\tau(\varphi)) \\ \tau(\varphi_1 \wedge \varphi_2) &= \text{seq}(\tau(\varphi_1), \tau(\varphi_2)) \\ \tau(\varphi_1 \vee \varphi_2) &= \text{first}(\tau(\varphi_1), \tau(\varphi_2)) \\ \tau(\varphi_1 \rightarrow \varphi_2) &= X \Rightarrow \text{seq}(\tau(\varphi_1), \text{first}(\text{stk} \Rightarrow X, \tau(\varphi_2)))@X\end{aligned}$$

$G \models \varphi$  if and only if  $\tau(\varphi)@G \longrightarrow^* G$

$G \not\models \varphi$  if and only if  $\tau(\varphi)@G \longrightarrow^* \text{stk}$

# Guarded systems

- define a new reduction relation

$$[K]_{\varphi} \Longrightarrow [K']_{\varphi} \text{ if } [K] \Rightarrow [K'] \text{ and } K' \models \varphi$$

- use strategies

$$[K]_{\varphi} \Longrightarrow \text{ifThenElse}(\tau(\varphi), X_1 \Rightarrow [K']_{\varphi}, X_2 \Rightarrow \text{error\_message})@K'$$
$$\text{if } [K] \Rightarrow [K']$$



# Conclusions (first part)

- port graphs: a biologically-inspired graphical structure
- biochemical calculus: a higher-order rule-based formalism
- verification of invariant properties
- applications to protein-protein interactions and autonomic systems

# Future work

- embed runtime verification
  - diagnose faults at execution and repair faults (adaptive behaviour)
  - identify properties to monitor
  - choose temporal logic: LTL<sub>3</sub> (T, ⊥, ?)
- add a stochastic semantics
- robustness analysis

# Abstractions for continuous-time Markov chains

# CTMCs

- state-based formalisms for describing dynamic systems:  $C = (S, s_0, R, L)$
- discrete steps, continuous time-steps
- suitable for modelling signalling pathways: stochastic, computational, concurrent

# CTMCs with levels

- population (species) based modelling
- discrete levels of concentrations
  - maximum molar concentration  $M$
  - choose granularity  $N$  for the abstraction, concentration step size  $H = M/N$
  - $0, 1, \dots, N$  levels of concentrations correspond to  $0, (0, H], (H, 2 * H], \dots, ((N-1) * H, N * H]$

# Formal model

- continuous time Markov chains with levels
- properties expressed as formulas in Continuous Stochastic Logic (CSL)
- symbolic probabilistic model checker PRISM

# Formal model

- mass-action kinetics
- reaction  $A + B \rightarrow C$  with  $k$  constant rate
- transition rate:  $k * (L_A * H) * (L_B * H) / H$

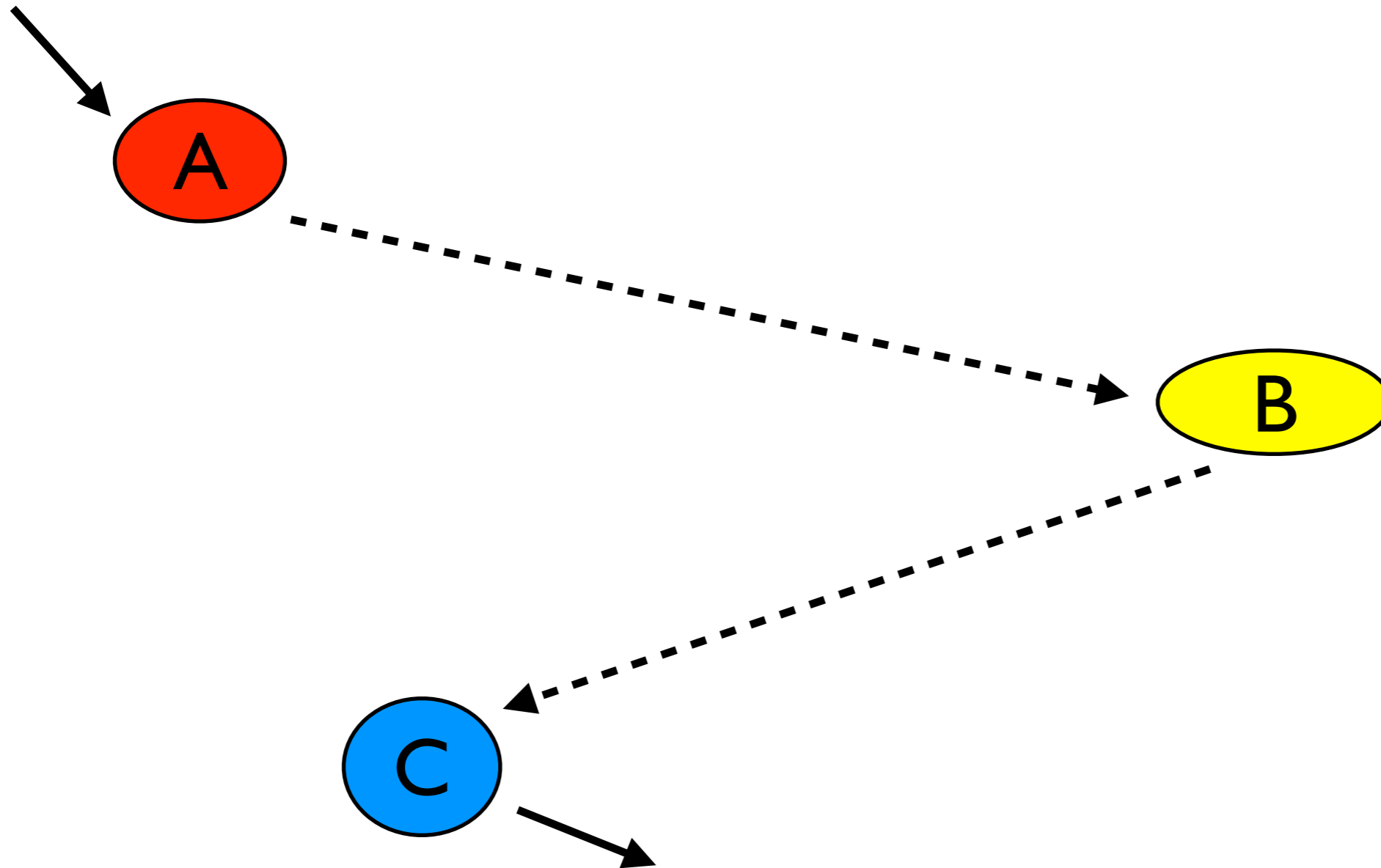
[rct1]  $L_A > 0 \rightarrow (L_A * H) : L_{A'} = L_A - 1$  // (in module for A)

[rct1]  $L_B > 0 \rightarrow (L_B * H) : L_{B'} = L_B - 1$  // (in module for B)

[rct1]  $L_C < \max_C \rightarrow 1 : L_{C'} = L_C + 1$  // (in module for C)

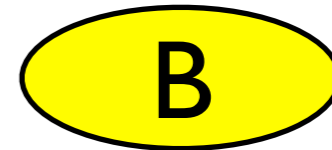
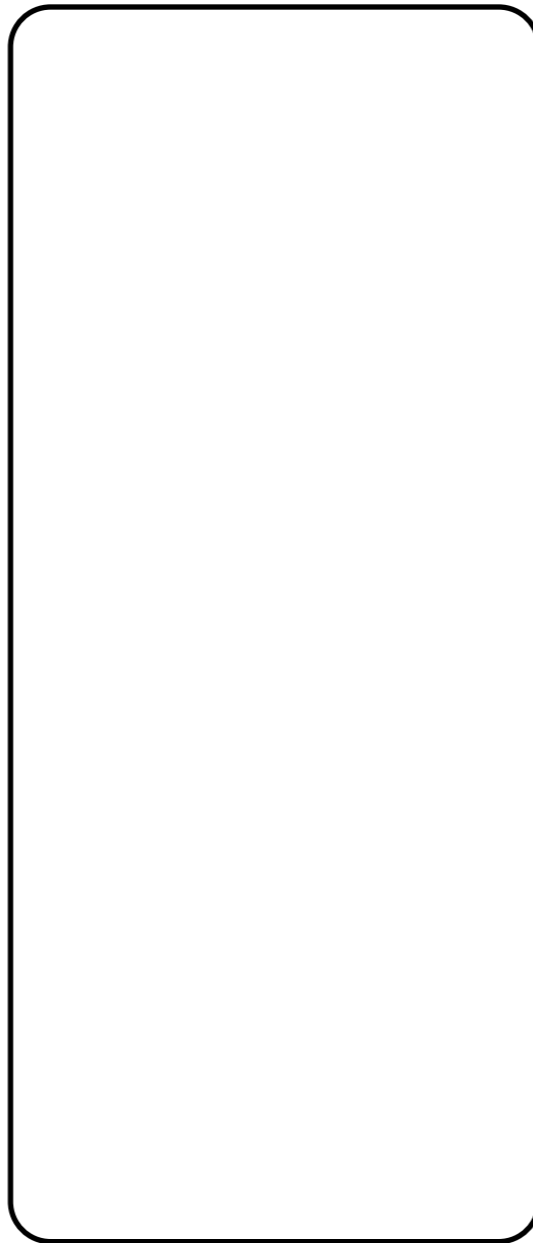
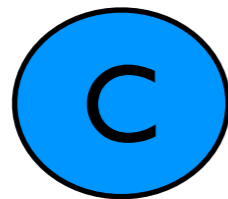
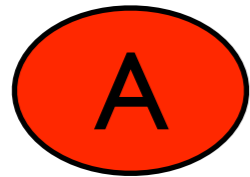
[rct1]  $\text{true} \rightarrow k/H : \text{true}$  // (in module for const)

# Signalling and scaffold proteins

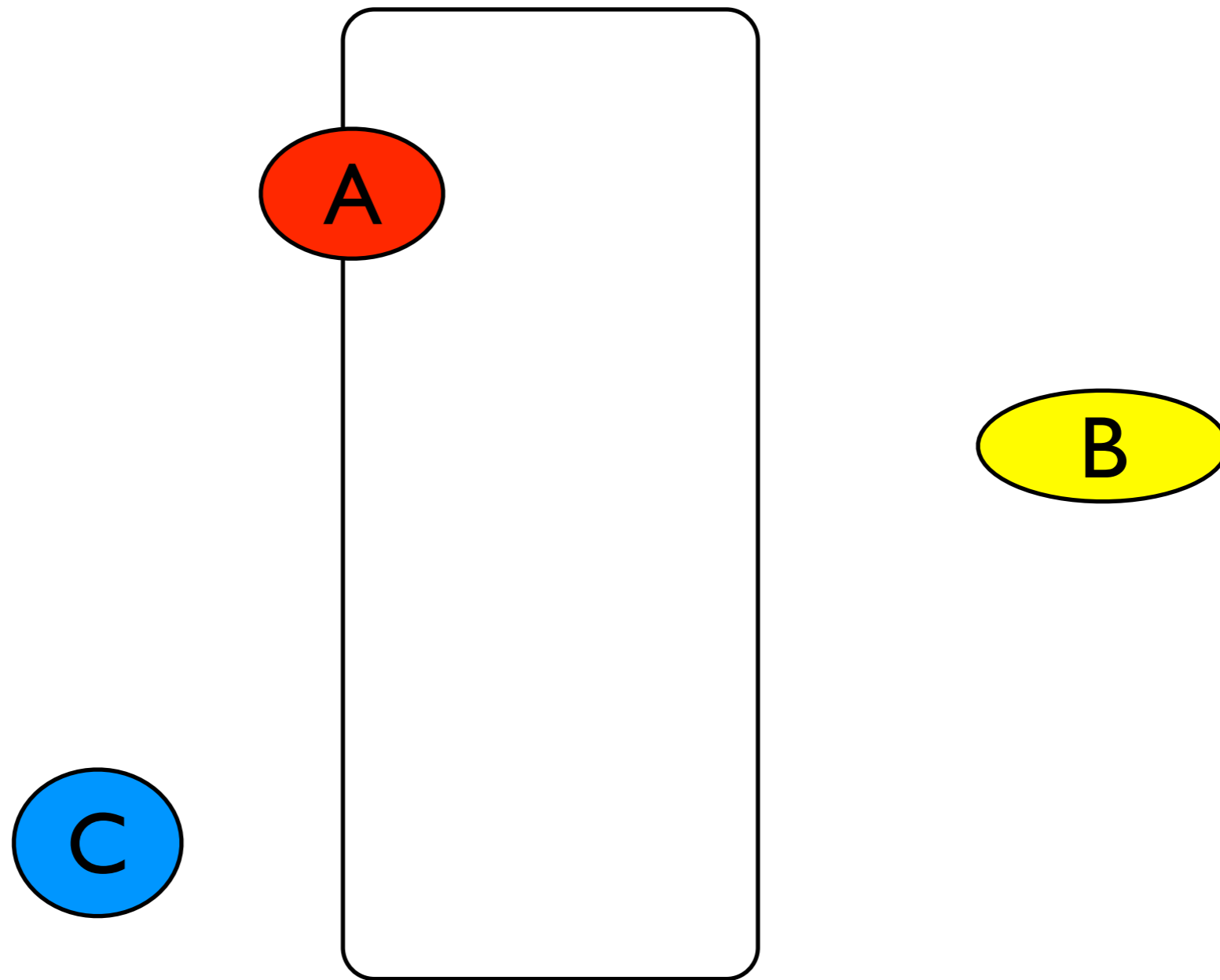




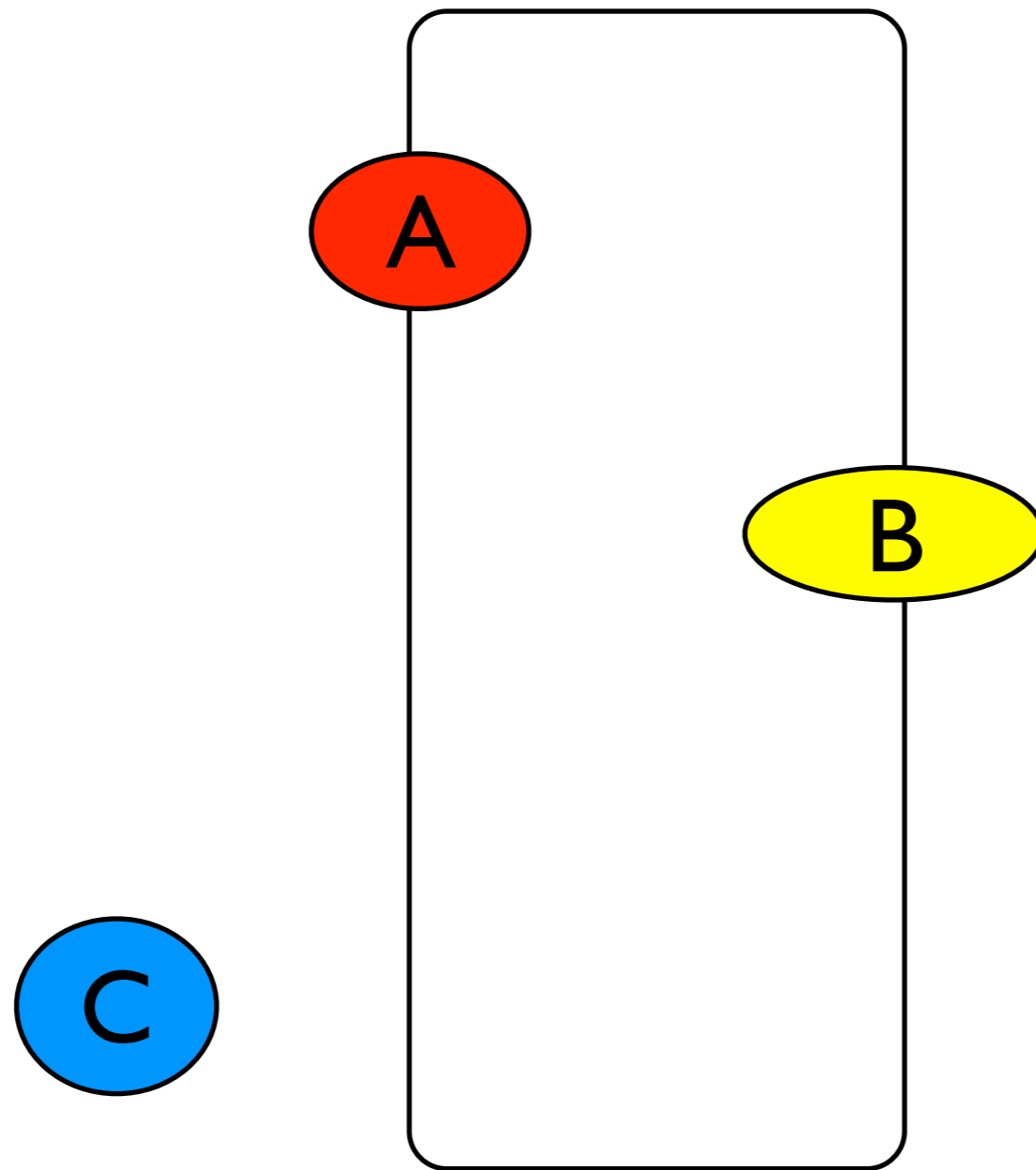
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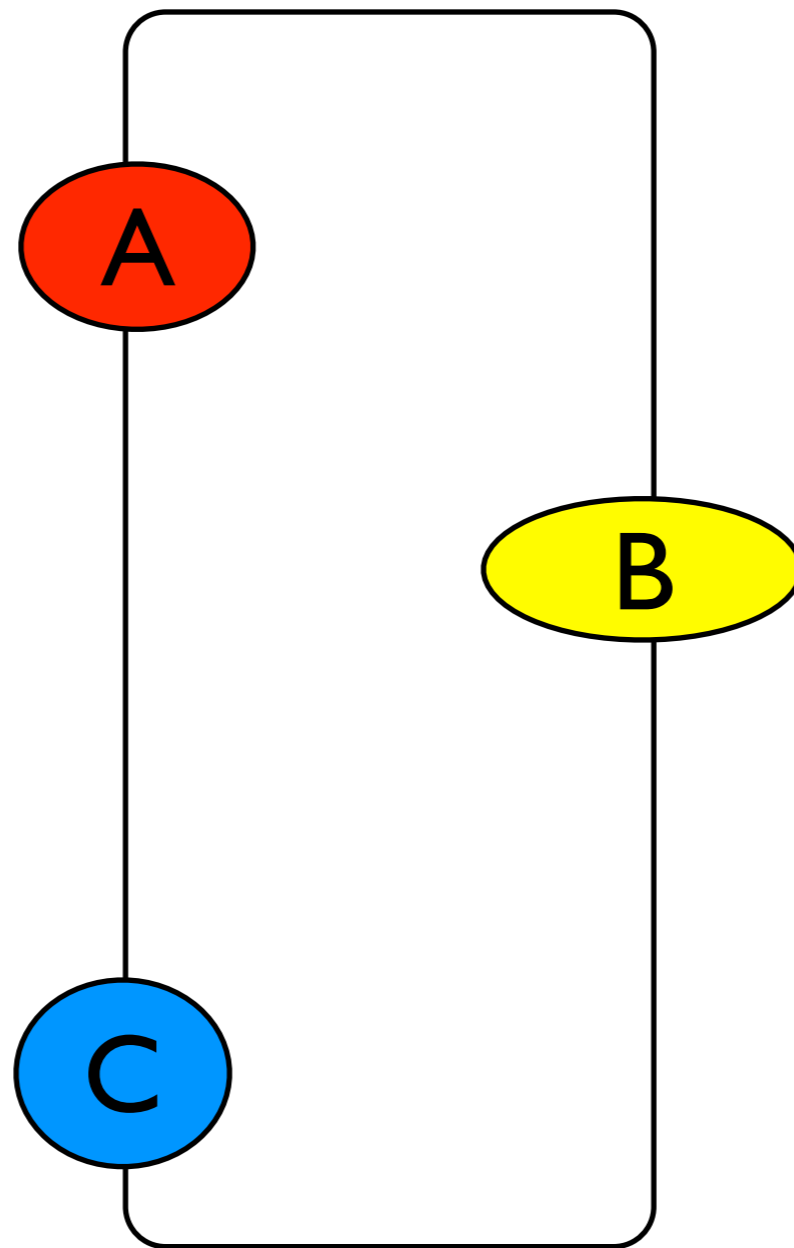
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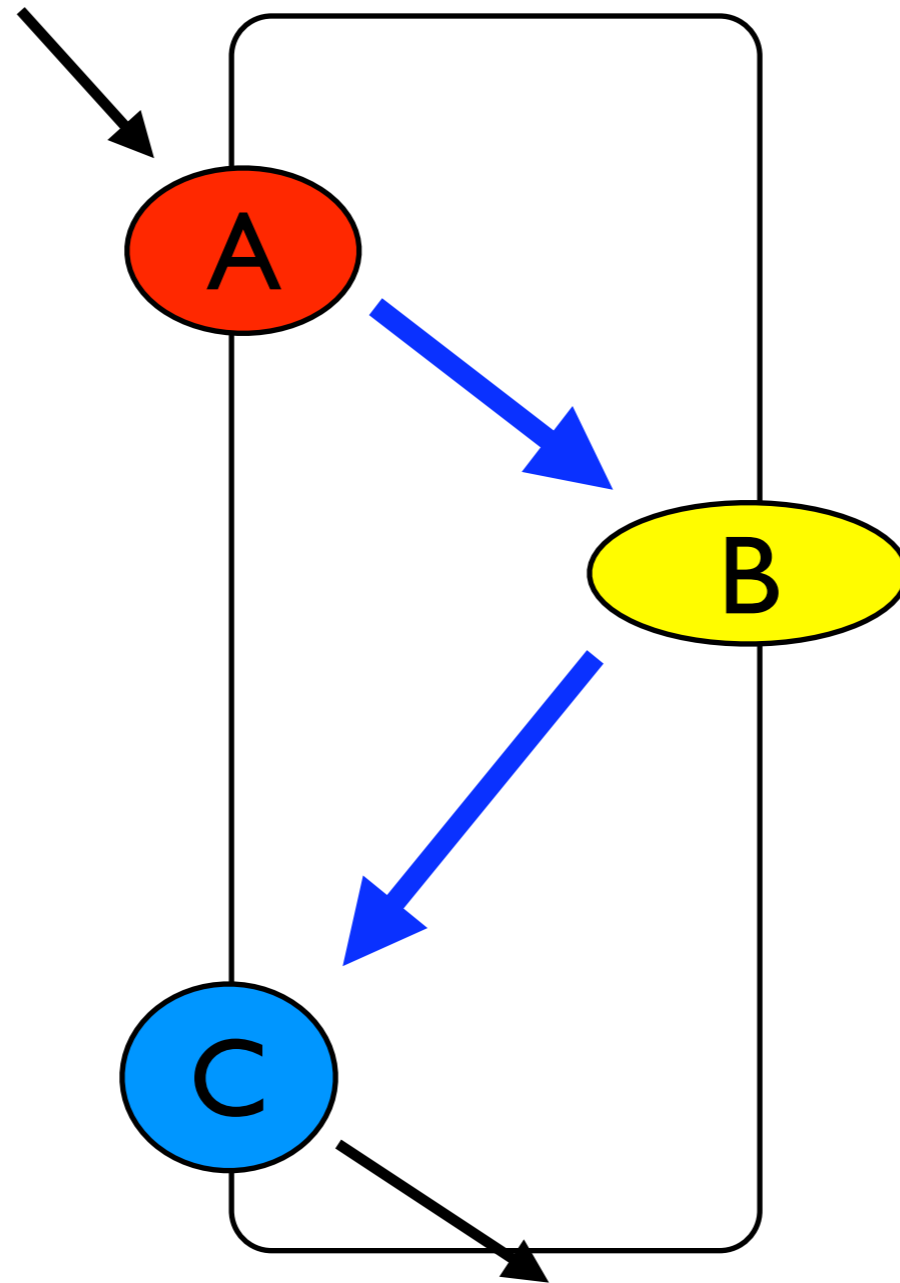
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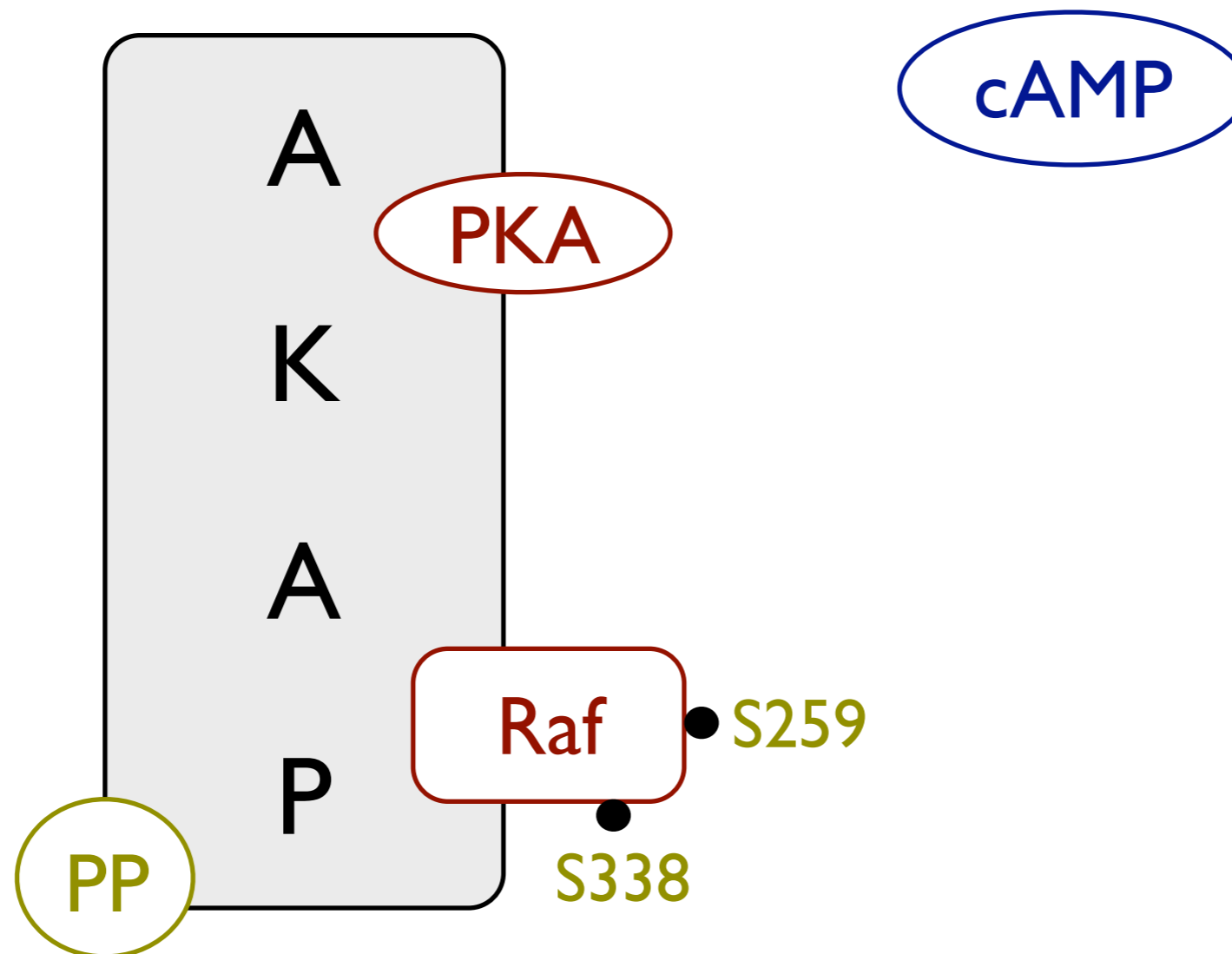


# Signalling and scaffold proteins



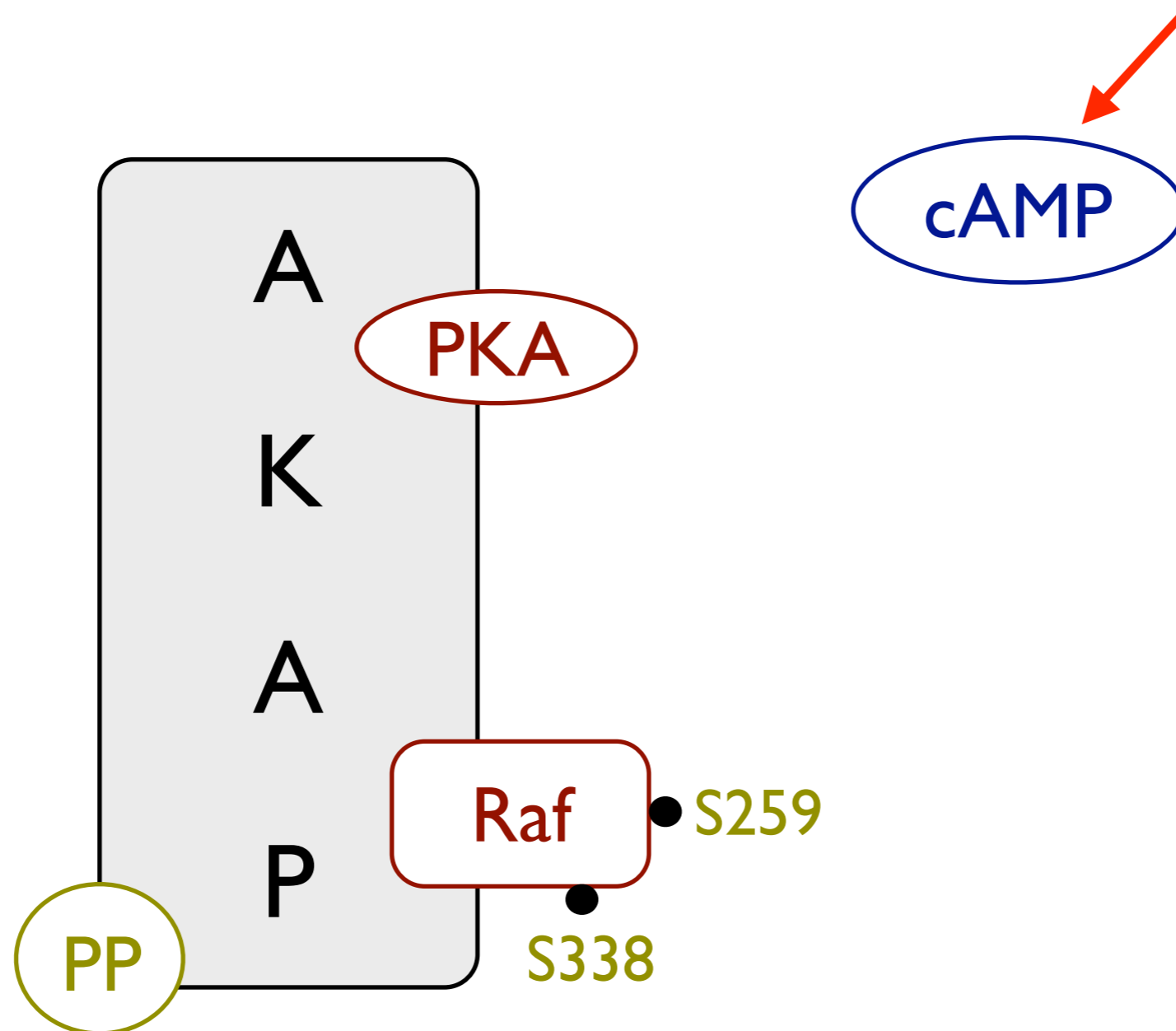
# AKAP

(A-kinase anchoring protein)



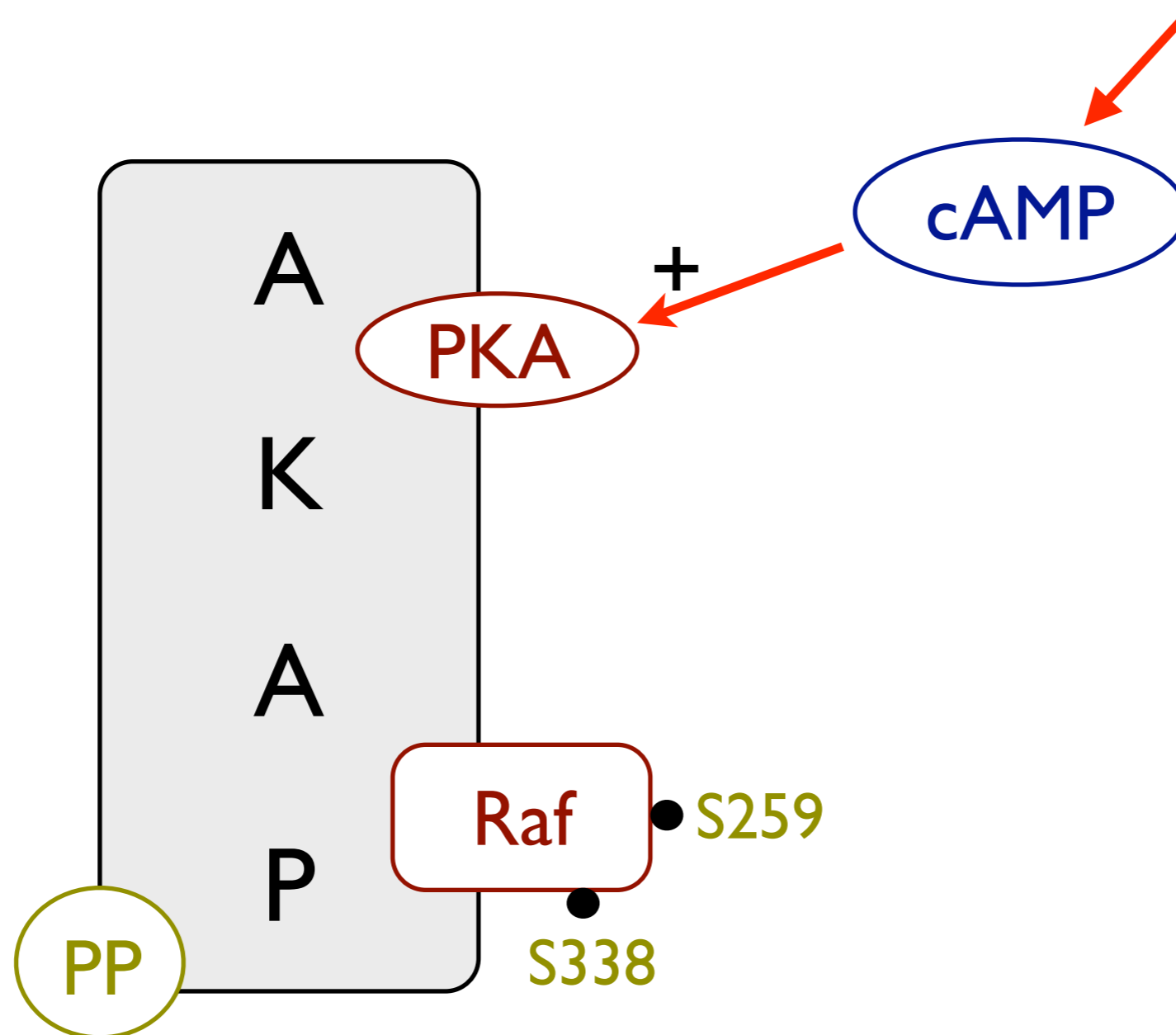
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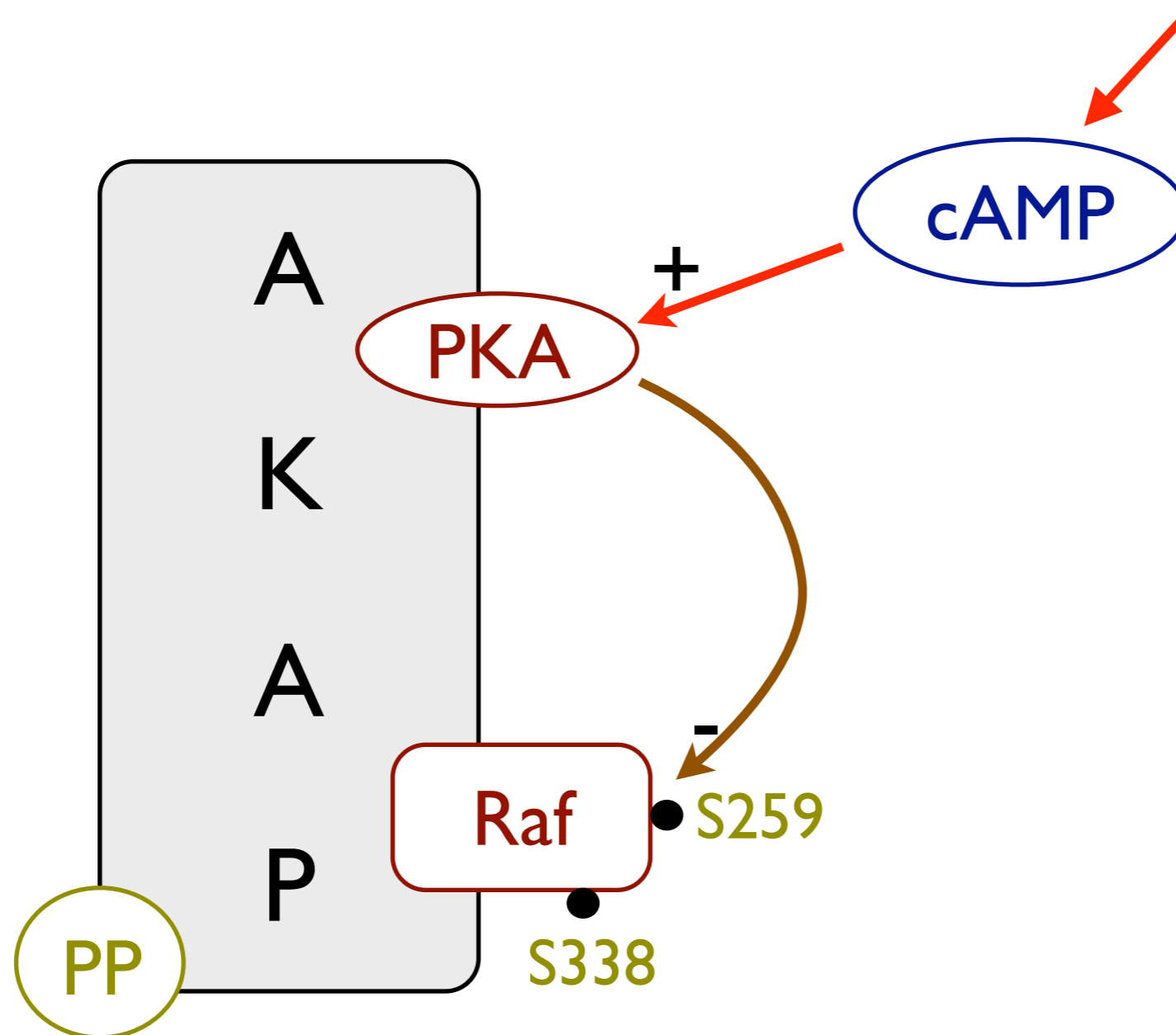
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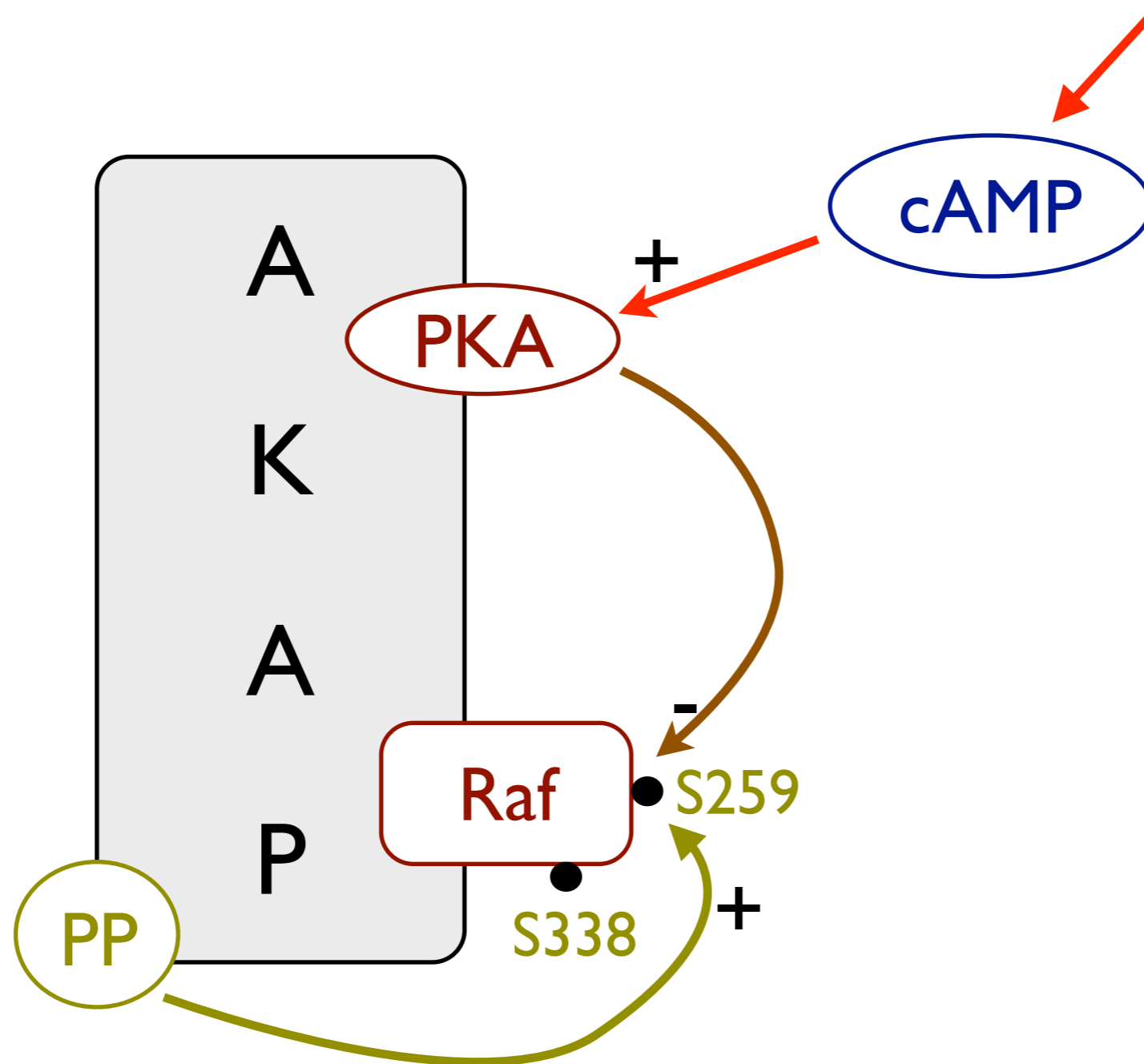
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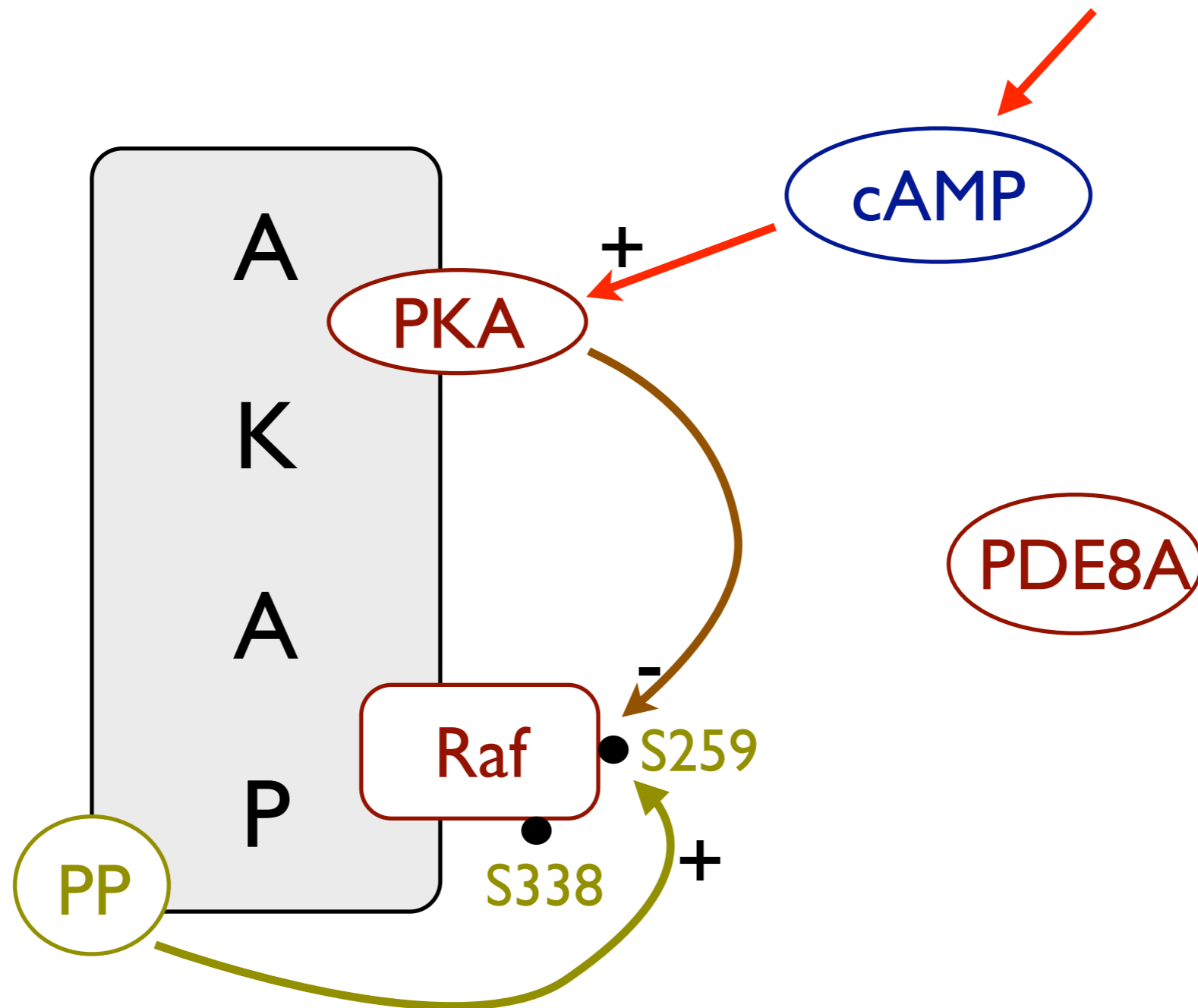
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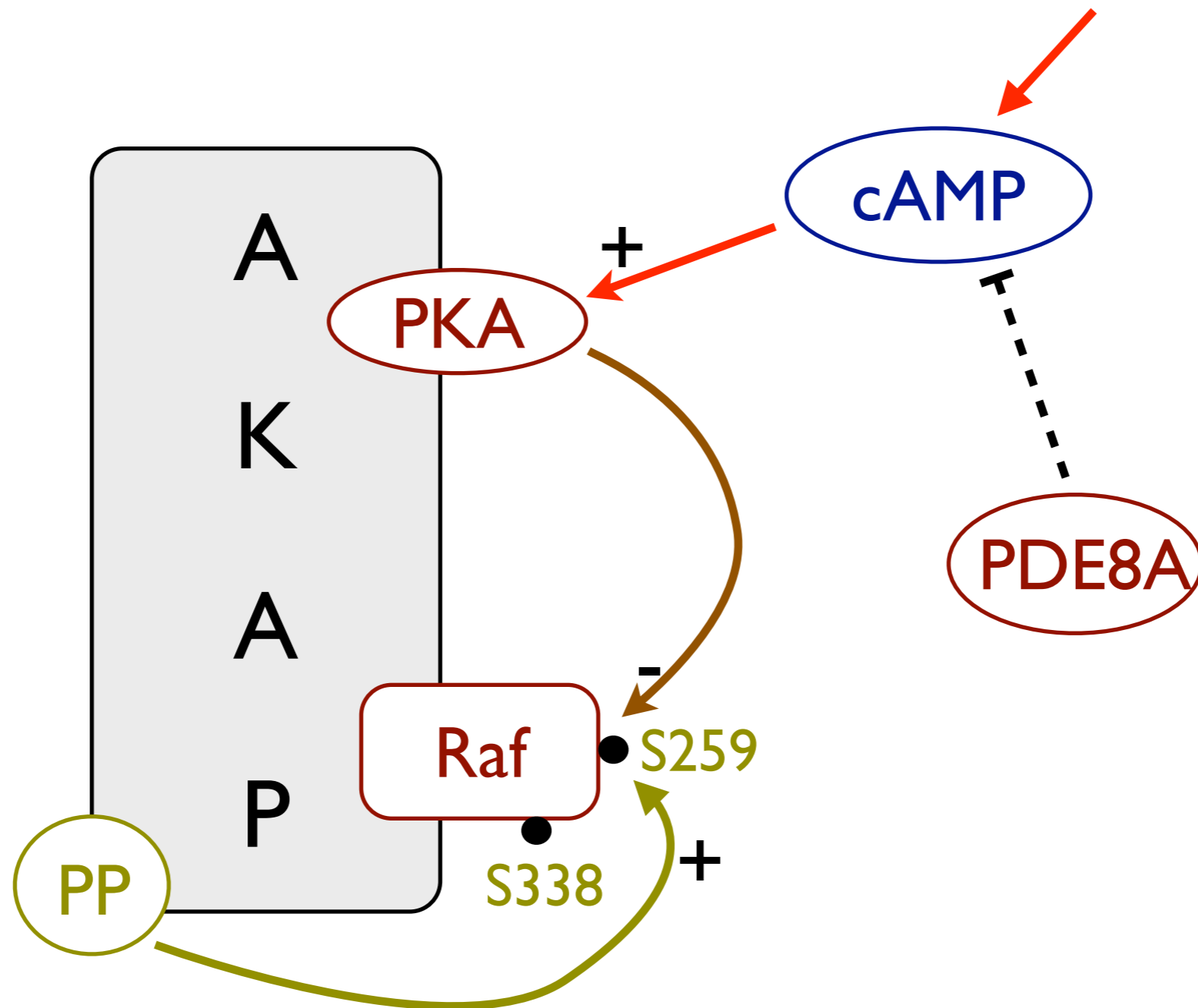
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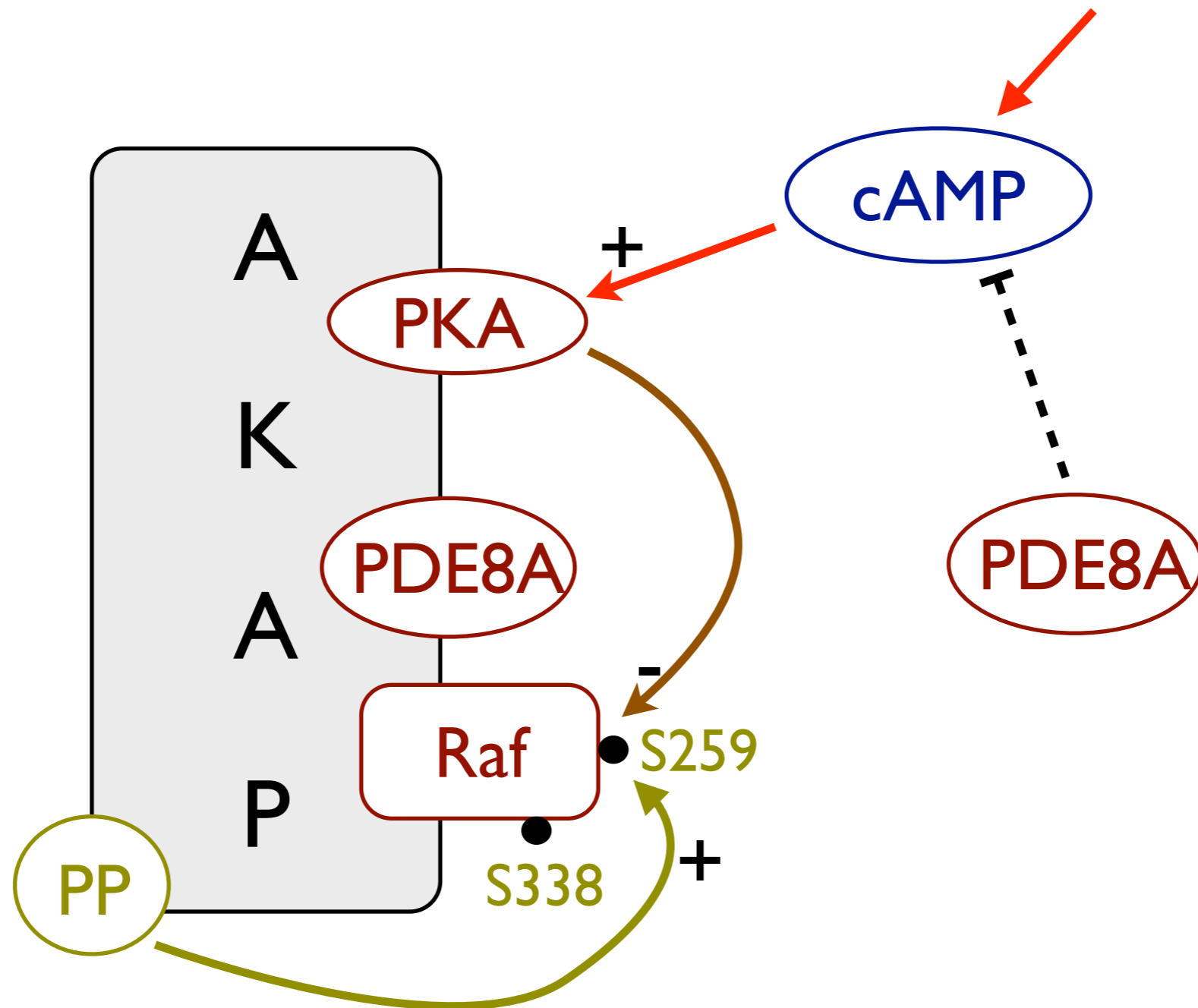
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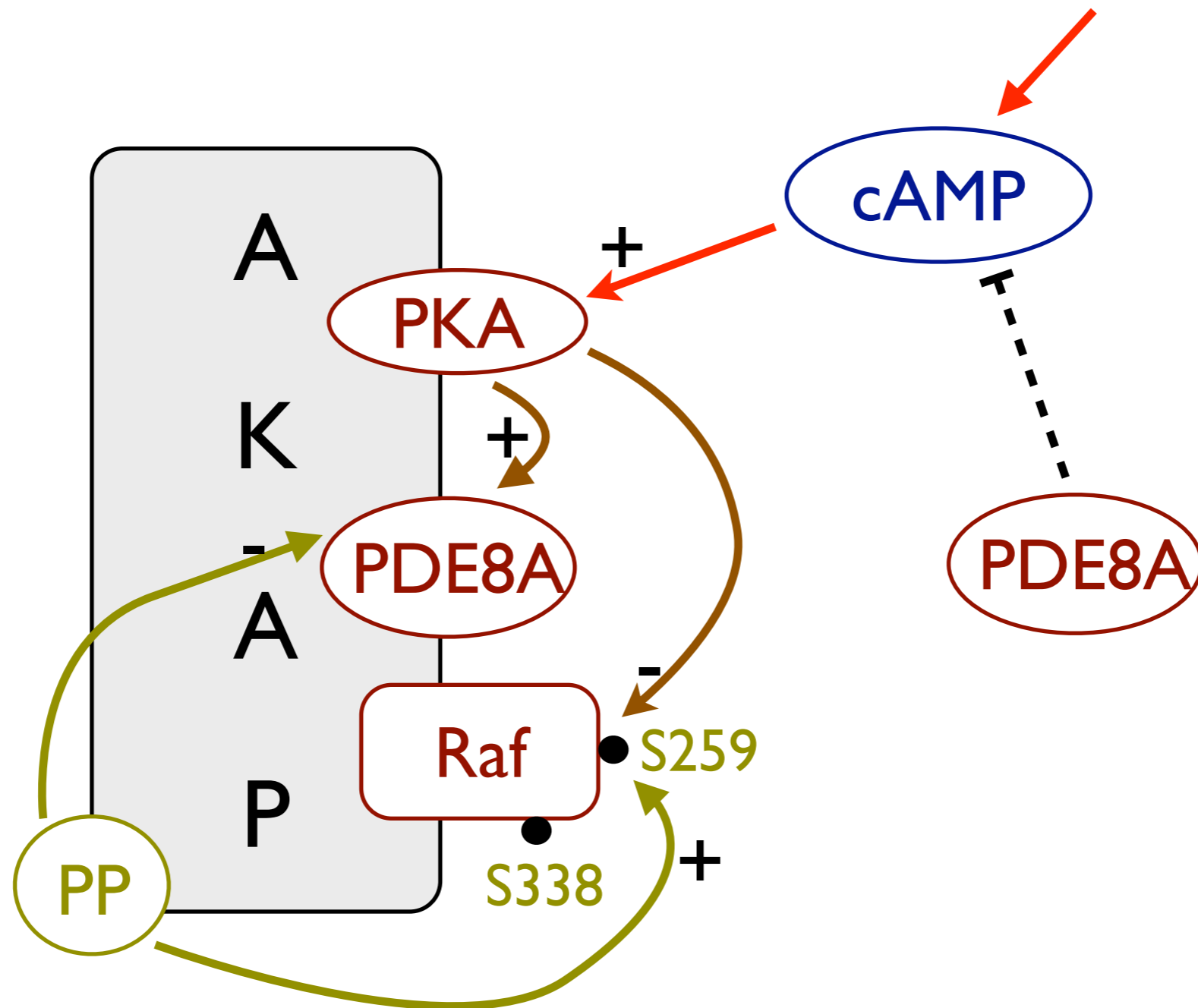
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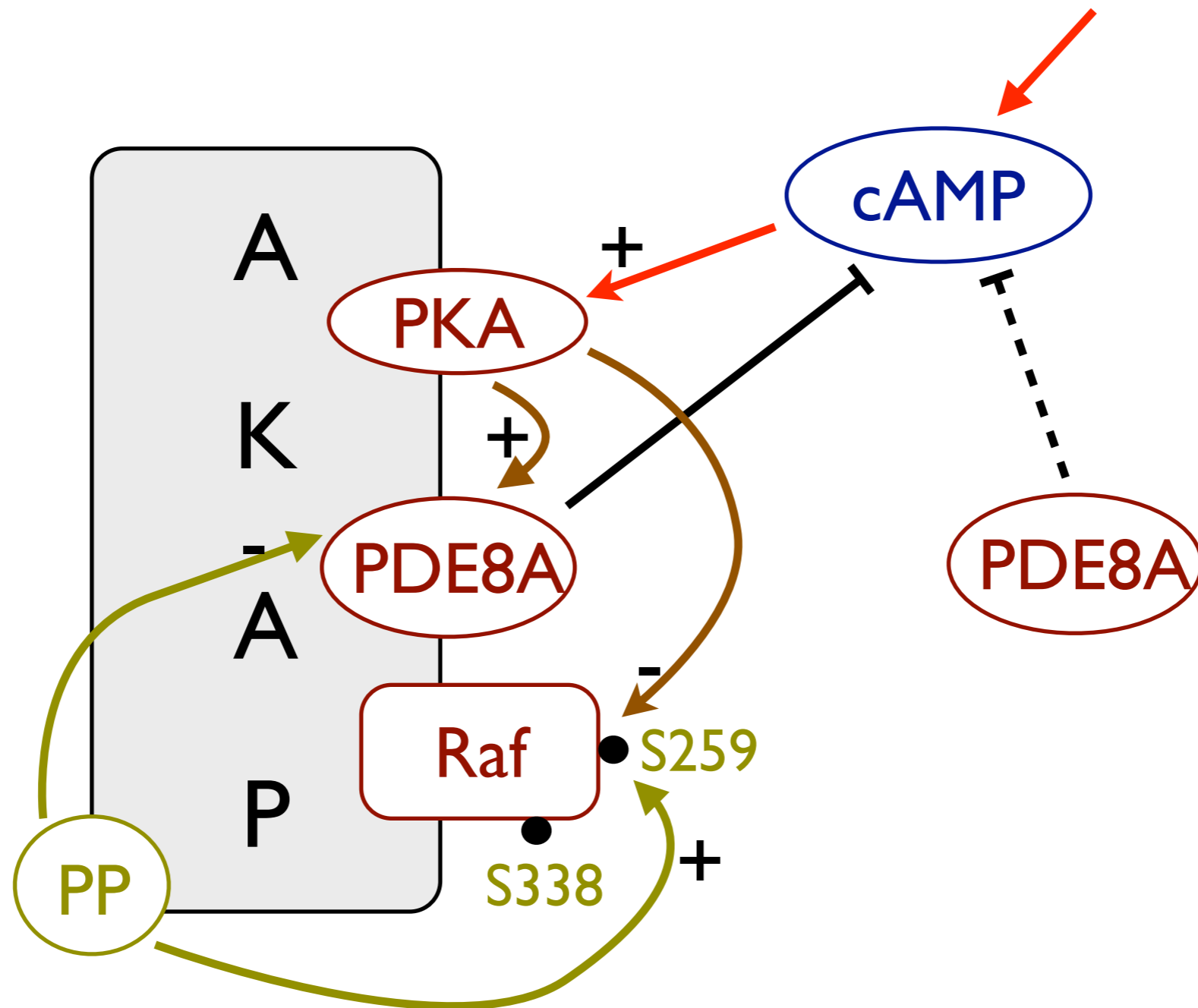
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# Expected behaviour

Q1:  $\uparrow$ pPDE8A I  $\Rightarrow$   $\downarrow$ cAMP  $\Rightarrow$   $\downarrow$ PKA<sup>+</sup>  $\Rightarrow$   $\uparrow$  Raf activity  
 $\Rightarrow$   $\downarrow$  pRafs259



# Expected behaviour

Q<sub>1</sub>: ↑ pPDE8A I ⇒ ↓ cAMP ⇒ ↓ PKA<sup>+</sup> ⇒ ↑ Raf activity  
⇒ ↓ pRaf<sub>S259</sub>

Q<sub>2</sub>: Pulsating behaviour

# PRISM model

- modules for cAMP, scaffold, free PDE8A I, PP
- mass action kinetics
- information on constant rates ratios

# Continuous Stochastic Logic

- extension of non-probabilistic CTL
- probability operator  $P$
- steady-state operator  $S$

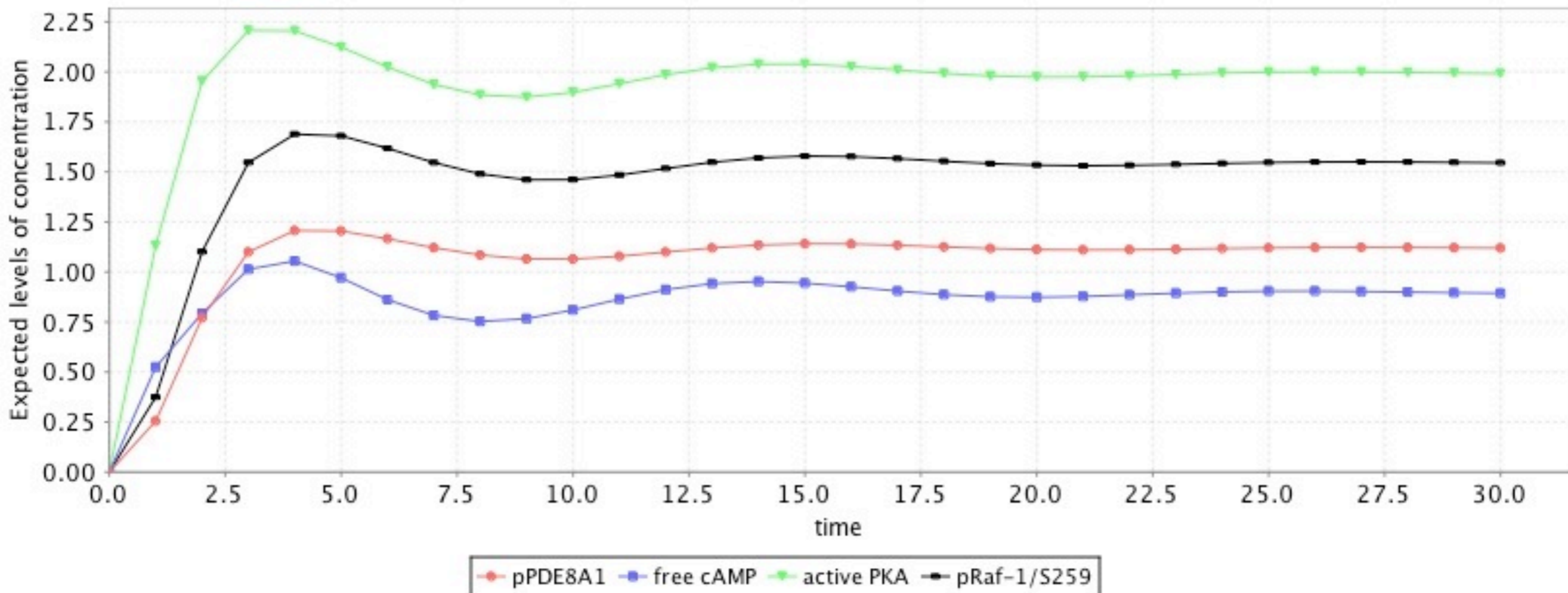
State formulae     $\Phi ::= \top \mid a \mid \neg\Phi \mid \Phi \wedge \Phi \mid P_{\bowtie p}[\phi] \mid S_{\bowtie p}[\Phi]$   
Path formulae     $\phi ::= X\Phi \mid \Phi U^I \Phi$

# Reward-based properties

- use of rewards (or costs) in CSL
  - real values assigned to states or transitions
  - to track variable values in states
  - to compute the expected value of a variable at a given time

# Reward-based properties

- state rewards for computing the expected levels for **cAMP**, **pPDE8A1**, **PKA<sup>+</sup>**, **pS259**



# Trend variables

- keep track of decreasing or increasing variable values
- define new variables in the PRISM modules:  
$$cAMP' = cAMP - 1 \quad \& \quad trend\_cAMP' = -1$$
- $\downarrow x$  ( $\uparrow x$ ) ascending (descending) trend for variable  $x$

# Necessarily preceded

[Monteiro et al. 08]

For  $\varphi = \downarrow \text{cAMP} \wedge \downarrow \text{PKA}^+$  and  $\psi = \uparrow \text{pPDE8A}$

CTL:  $(\text{EF } \varphi) \wedge \text{AG}((\neg \psi) \Rightarrow \text{AG}(\neg \varphi))$

CSL:  $P_{>0}[\text{F } \varphi] \wedge P_{\leq 0}[\text{F}(\neg((\neg \psi) \Rightarrow P_{\geq 1}[\text{F}(\neg \varphi)]))]$

# Pulsations

Show that the levels of pPDE8A I fluctuate:

- $\varphi = \uparrow_{\text{pPDE8A I}}$  and  $\psi = \downarrow_{\text{pPDE8A I}}$
- pulsation in CTL [Fages05, Ballarini et al. 09]:

$$\text{AG}((\varphi \Rightarrow \text{EF}\psi) \wedge (\psi \Rightarrow \text{EF}\varphi))$$

- pulsation in CSL:

$$P_{\leq 0}[F (\neg(\varphi \Rightarrow P_{>0}[F\psi]) \vee \neg(\psi \Rightarrow P_{>0}[F\varphi]))]$$



# Pulsations

- for cAMP:  $\varphi = \uparrow \text{cAMP}$  and  $\psi = \downarrow \text{cAMP}$
- for PKA<sup>+</sup>:  $\varphi = \uparrow \text{PKA}^+$  and  $\psi = \downarrow \text{PKA}^+$
- coordinated pulsations:

$$\varphi = \uparrow \text{pPDE8A} \wedge \downarrow \text{cAMP} \wedge \downarrow \text{PKA}^+ \text{ and}$$

$$\psi = \downarrow \text{pPDE8A} \wedge \uparrow \text{cAMP} \wedge \uparrow \text{PKA}^+$$

# Overview of AKAP modelling

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- formal model of a biological process
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- refine the model with more experimental data
- trend variables, amplitude of oscillations
- formulate new properties and express them using a temporal logic

# Abstractions for CTMCs with levels

- relation between two CTMCs with levels for the same system:
- aim: preserve temporal properties and do model checking on the more abstract model
- if  $C^N \models \varphi$ , then  $C^{kN} \models f(\varphi)$  - who is  $f$  ?
- (weak) simulation relation [Baier et al.] does not work...



# Temporal properties

- classification of temporal properties for signalling pathways
- BIOCHAM [Fages et al.]
- patterns [Monteiro et al.08]
- stochastic models, not only qualitative or probabilistic

# Temporal properties

- is CSL expressive enough?
- what about LTL(R) ? [Fages et al.]
- linear versus branching time for biologists?
- satisfaction probabilities for biologists?

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**Thank you!**  
**Questions?**

# Bibliography

- **[Monteiro et al. 08]** Pedro T. Monteiro, Delphine Ropers, Radu Mateescu, Ana T. Freitas, and Hidde de Jong. *Temporal logic patterns for querying dynamic models of cellular interaction networks*. *Bioinformatics*, 24(16):227--233, 2008.
- **[Ballarini et al. 09]** Paolo Ballarini, Radu Mardare, and Ivan Mura. *Analysing Biochemical Oscillation through Probabilistic Model Checking*. *ENTCS*, 229(1):3--19, 2009
- **[Fages05]** François Fages. *Temporal Logic Constraints in the Biochemical Abstract Machine BIOCHAM*. *LOPSTR'05*, volume 3901 of *LNCS*, pages 1--5. Springer, 2005.